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Nervous system development of a primitive pulmonate (Mollusca: Gastropoda) and its bearing on comparative embryology of the gastropod nervous system

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ABSTRACT As is typical for primitive pulmonates, the ellobiid *Ovatella myosotis* undergoes a veliger stage during development. The development of the nervous system and sensory organs of this species was examined in detail, using 2 µm serial sections. During the larval phase, three pairs of ganglia in the headfoot (cerebropleural, pedal and buccal ganglia) and three visceral loop ganglia (supraoesophageal, suboesophageal and visceral ganglion) are formed. The first-formed connective from the cerebropleural ganglion to the pedal ganglion is the pleuropedal connective. The pleural ganglia separate from the cerebral ganglia after metamorphosis. Eyes and statocysts are formed by ectodermal invaginations. An osphradial ganglion (the remnant of an osphradium) is formed during the larval phase and reduced at metamorphosis. A detailed discussion on nervous system genesis in gastropods is provided. The development of *O. myosotis* reveals many similarities in the nervous system genesis of pulmonates and opisthobranchs. This allows a critical evaluation of recent studies on other euthyneuran nervous system genesis. The sequence of formation of connectives to the pedal ganglion in *O. myosotis* provides valuable information for an interpretation of these structures in opisthobranch larval nervous systems. This throws light on the question of the position of the pleural ganglion in nudibranchs. Also, it is suggested that the formation of the pleural ganglion from a common anlage (= rudiment) with the cerebral ganglion is typical for higher gastropods.

RIASSUNTO L'ellobiide *Ovatella myosotis* possiede uno stadio di veliger durante il suo sviluppo, come è tipico per i pulmonati primitivi. Lo sviluppo del sistema nervoso e degli organi di senso è stato esaminato in dettaglio attraverso sezioni seriali di 2 µm. Durante la fase larvale si formano tre paia di gangli (cerebropleurale, pedale e gangli buccali) nel complesso cefalopodiale, e tre gangli viscerali (supraesofageo, subesofageo e viscerale). Il primo connettivo a formarsi dal ganglio cerebropleurale a quello pedale è il connettivo pleuropedale. I gangli pleurali si separano dai gangli cerebrali dopo la metamorfosi. Gli occhi e le statocisti si formano per invaginazione ectodermica. Un ganglio osfradiale (il residuo di un osfradio) si forma durante la fase larvale e si riduce alla metamorfosi. Si discute dettagliatamente sulla genesi del sistema nervoso nei gasteropodi. Lo sviluppo di *O. myosotis* rivela molte similarità nella genesi del sistema nervoso con pulmonati ed opistobranchi. Questo permette una valutazione critica di studi recenti sulla genesi del sistema nervoso di altri eutineuri. La sequenza di formazione dei connettivi al ganglio pedale in *O. myosotis* fornisce importanti informazioni per una interpretazione di queste strutture nel sistema nervoso larvale degli opistobranchi. Ciò fa luce sulla questione della posizione del ganglio pleurale nei nudibranchi. Inoltre, si suggerisce che la formazione del ganglio pleurale da un *anlage* (= rudimento) comune con il ganglio cerebrale, sia tipica per i gasteropodi superiori.

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INTRODUCTION

Increasing attention has been paid to anatomical characters of the nervous system in more recent analyses of molluscan and gastropod phylogenetic relationships (HASZPRUNAR, 1988; NORDSIEK, 1992; HUBER, 1993). The selected characters originate from anatomical studies of adult animals as well as from embryological studies. Embryology was called upon to interpret homology relations of adult structures and was the topic of a number of recent publications. This research, which focused mainly on nudibranch opisthobranchs, led to conclusions extending beyond the systematic level of gastropods (PAGE, 1992a,b) or even molluscs (PAGE, 1993). In contrast to opisthobranchs, the pulmonates have hardly received any attention during the last few decades in this regard.

The data on pulmonates are unsatisfactory for two reasons: (1) as opisthobranch studies show, modern techniques provide a substantially more detailed data set and demonstrate the limited reliability of embryological studies carried out with the paraffin section technique; (2) all previous studies on the development of pulmonate nervous systems were restricted to species with direct development, i. e., those lacking the typical veliger larval stage. Direct development while common among pulmonates, is nevertheless a derived condition as in all major gastropod taxa indirect development via a veliger larva is the primitive condition. In direct development the shape of developmental stages and the timing of structure formation are usually substantially altered compared with primitive larval development and therefore the usefulness of species with a suppressed larval stage for comparing developmental processes between higher gastropod taxa is limited.

The ellobiid pulmonate *Ovatella myosotis* (Draparnaud, 1801) has, as is typical for primitive pulmonates, a veliger larva. This larval stage is similar to the primitive planktotrophic euthyneuran larva, although in this species the larval phase is spent inside the egg capsule. *O. myosotis* is very easy to culture and therefore convenient for embryological studies. It is particularly appropriate for developmental studies of the nervous system because the major ganglia in the adults are separated. Consequently, it is possible to interpret nervous system elements of developmental stages by tracing their genesis in reverse direction (form late to early stage). This should be carried out with a method that allows the investigation of developmental processes as continuously as possible. Until now most studies on gastropod nervous system development did not do this sufficiently and described the organisation of more or less isolated developmental stages only.

Contradictory interpretations on the larval nervous system of nudibranch opisthobranchs have recently been published (CARROLL & KEMPF, 1994 vs. PAGE, 1992a,b vs. TARDY, 1970, 1974). Adult nudibranchs have an extremely concentrated arrangement of ganglia. It is surprising that representatives of this group were chosen for recent studies because tracing and interpreting larval elements is extremely difficult if the respective elements are not separate in the adult. The development of *O. myosotis* may provide valuable information on the identity of structures of larval pulmonate and even opisthobranch nervous systems. According to Von Baer's rule (see GOULD, 1977, DENIS & COLLENOT, 1995a,b) the earlier the developmental stages of different species are, the more similar they are. This is also applicable to larvae of euthyneurous gastropods. The available studies on pulmonates with veliger development (FRETTER, 1943; LITTLE *et al.*, 1985) have not revealed any character generally different from that of opisthobranchs and as will be shown, the larval nervous system of pulmonates closely resembles that of opisthobranchs.

The present study may also be useful as basis for future investigations with modern techniques like histochemistry or immunocytochemistry in this species. The knowledge of the general nervous system morphology would be very helpful to carry out studies like RAINERI (1995) or KEMPF et al. (1997) did in other molluscs.

The development of the sensory organs is included in this study because these organs often provide significant information about the nervous system.

MATERIAL AND METHODS

Adult specimens of *Ovatella myosotis* were collected near Alberoni on the Lido di Venezia, Italy. They were cultured in porous flower pots (upper diameter: 24 cm) whose bottoms were filled with a 3-cm layer of substratum taken from the collection site and further covered with coarse potsherds. The bowls were placed in saucers filled with seawater in order to regulate substratum moisture and salinity. The salinity in the saucer was kept between 15 and 35 per mille and the pots were covered with plastic foil to maintain high humidity inside. The snails were fed with standard aquarium fish food. This set up provided a self-reproducing population over many years.

The animals deposited egg masses on the underside of the potsherds or in shallow depressions in the substratum. The egg masses were isolated as follows: Shallow holes (depth: 5 mm; width: 4 mm) were drilled into a piece of ceramic tile. Single egg masses were placed inside these holes. This tile was then covered with a second flat ceramic piece. The entire structure was placed inside the culture bowl at the same level where egg masses were originally deposited. This was the only satisfactory method of isolating egg masses because, in particular, early stages showed a high rate of anomalies and mortality if removed from the culture bowls.

For histological investigation samples of 10 - 15 eggs were taken from isolated egg masses on three to five subsequent days. Specimens were freed from the egg capsule with the help of minute insect needles. Developmental stages younger than six days were fixed in 4 % seawater-buffered formalin before removal from the egg capsule to minimise damage. Older stages were freed from the egg capsule in seawater and subsequently fixed in Bouin's fixativ, which showed better histological results. Developmental stages capable of movement and retraction were anaesthetised prior to removal. For this purpose, samples were put for one to two hours, in a seawater-filled dish to which a drop of ethylene-dissolved menthol was added. Hatched juveniles and adults were anaesthetised in equal parts of 7.5% $MgCl_2$ and seawater or with menthol as described above. Hatched animals were starved for at least 24 hours to clear the gut of sand particles which could negatively effect sectioning. These animals were fixed with Bouin's or Susa's fixative fluid.

Dehydration was carried out in a graded series of ethanol. Specimens up to a size of 3.5 mm were transferred into propylene oxide and embedded in araldite (MOLLENHAUER, 1964). They were sectioned on a Reichert Autocut microtome with ralph glass knives (BENNETT *et al.*, 1976), which were broken on a Reichert ralph knife maker. Two micrometer sections were ribboned with a technique modified from HENRY (1977). A layer of contact cement (Pattex compact, Henkel Co., Vienna) was put on the underside of the block. Staining was done with Regaud's iron hematoxylin or Richardson's methylene-blue - azur II. Larger specimens were transferred to methylbenzoate and benzene and embedded in paraplast. These five or seven µm sections were stained in Heidenhain's azan.

The age of the intracapsular developmental stage was determined as follows: Egg masses were divided into samples which were fixed on different - usually three to five subsequent - days. These sample series covered the entire intracapsular development. The absolute age of each sample was determined by equating identical stages of overlapping sample series. This was controlled by comparing stages with known time of egg deposition.

Four to ten serially sectioned specimens of each day of the intracapsular development period and of a number of hatching stages, juveniles and adults were examined. This high number of investigated specimens enabled the genesis of structures to be followed in a nearly continuos way. Nervous system reconstructions were prepared of specimens at day 5 - 6, 8, 10 - 13, 15, 19, of a hatching stage and of an adult animal. Serial sections were reconstructed by measuring distances on sections with an ocular micrometer on a Reichert Biovar microscope. Contours of resin embedded specimens were established by drawing the embedded specimen with a camera lucida or by measuring distances using the edge of the plastic section as reference. For reconstruction of the adult nervous system, distances of nervous elements to pedal sole and foregut were measured. Photographs were taken on a Reichert Diavar microscope.

RESULTS:

(a) Adult anatomy:

The following anatomical data and terminology are important in understanding the developmental processes and the reasoning behind the discussion in this study.

The general organisation of the adult nervous system of *Ovatella myosotis* is given in figure 1. In the adult animal the cerebral ganglia are strongly asymmetrically arranged because the penis complex causes the right ganglion to lie more posteriorly and ventrally than the left one (Fig. 1). The procerebrum (Fig. 1, PR) is an anteriorly separated portion of the cerebral ganglion, which is innervated by two nervous junctions, the anterior and posterior procerebral connectives. These connectives are positioned next to the anterior and posterior ends of the procerebrum. The procerebrum is the base of the tentacular nerve (Fig. 1, TN), which splits into three branches in some distance from the procerebrum. Fibers of the optic nerve (Fig. 1, ON), which emerges near the tentacular nerve, can be traced to the small posterior procerebral connective, which is formed by these fibers. The cerebral gland (Fig. 1, CG) is consisting of a remarkable structure, a duct with a tiny lumen leading from the procerebrum to the body surface at the lateral base of the tentacle, where it opens with a pore. The cerebral commissure (Fig. 1, CC), the connectives to the ganglia, and all other nerves emerge from the main portion of the cerebral ganglion. There are three anteroventrally emerging labial nerves on the left side and two on the right side (Fig. 1, L1 - L3). The penis nerve (Fig. 1, PSN) emerging from the right cerebral ganglion corresponds to one labial nerve from the left side (Fig. 1, L1). The statocyst nerve (Fig. 1, SN) emerges posteriorly between the cerebropedal and the cerebropleural connective. Dorsally the cerebral ganglion is covered with a cap of presumably neurosecretory tissue (Van Mol, 1967), the dorsal body (Fig. 1, D).

The pedal ganglia of *Ovatella myosotis* have three ventrally (Fig. 1, VP1 - VP3) and three laterally (Fig. 1, LP1 - LP3) emerging nerves. In addition to the pedal commissure, the pedal ganglia are connected by the parapedal commissure, which emerges from the posterior portion of the ganglia.

The statocysts lie embedded in the pedal ganglia (Fig. 1) adjacent to the origin of the pleuropedal connective. The connectives are lateroanterior to the statocysts. Near its entry into the statocyst, the statocyst nerve is twisted lateroposteriorly around the pleuropedal connective in posthatching juveniles and adults. In this area both nerves are in intimate contact with each other and near the statocysts, the fibers of the statocyst nerve are hardly distinguishable from those of the pleuropedal connective.

The pleural ganglia (Fig. 1, PL) lie very close to the pedal ganglia (hypoathroid condition) and have no nerves. The suboesophageal ganglion (Fig. 1, SBG) gives rise to one dorsal nerve (Fig. 1, SBN). A short distance from the suboesophageal gan-

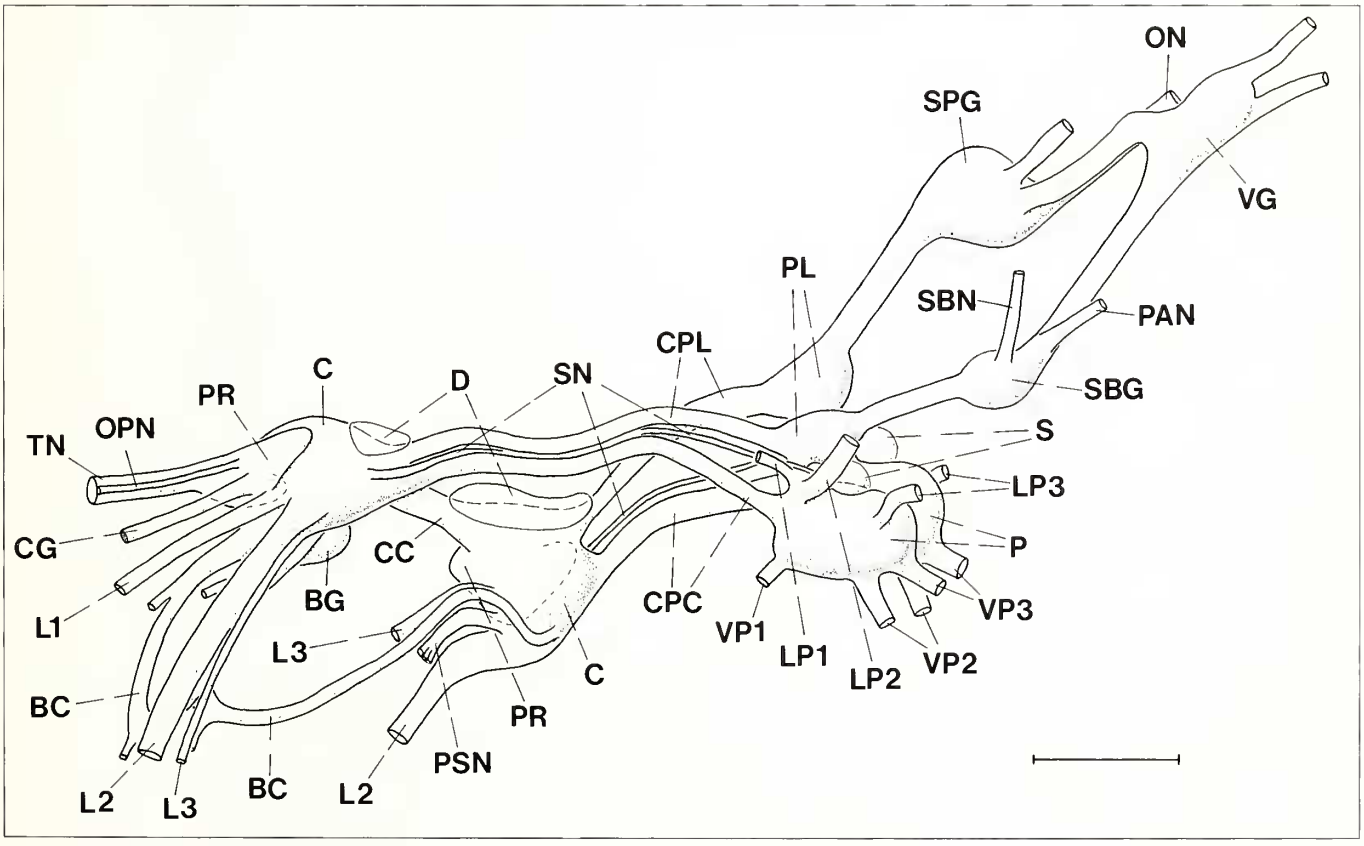

Fig. 1. Reconstruction of the nervous system of an adult specimen of *Ovatella myosotis*. Lateral view from the left side. BC = buccal connective; BG = buccal ganglion; C = cerebral ganglion; CC = cerebral commissure; CG = cerebral gland; CPC = cerebropedal connective; CPL = cerebropleural connective; D = dorsal body; L1-3 = labial nerves; LP 1-3 = lateral pedal nerves; ON = osphradial nerve; OPN = optic nerve; P = pedal ganglion; PL = pleural ganglion; PR = Procerebrum; PSN = penis nerve; S = statocyst; SBG = suboesophageal ganglion; SBN = suboesophageal ganglion nerve; SN = statocyst nerve; SPG = supraoesophageal ganglion; TN = tentacle nerve; VG = visceral ganglion; PAN = pallial nerve; VP 1-3 = ventral pedal nerves. Scale bar = 40 µm

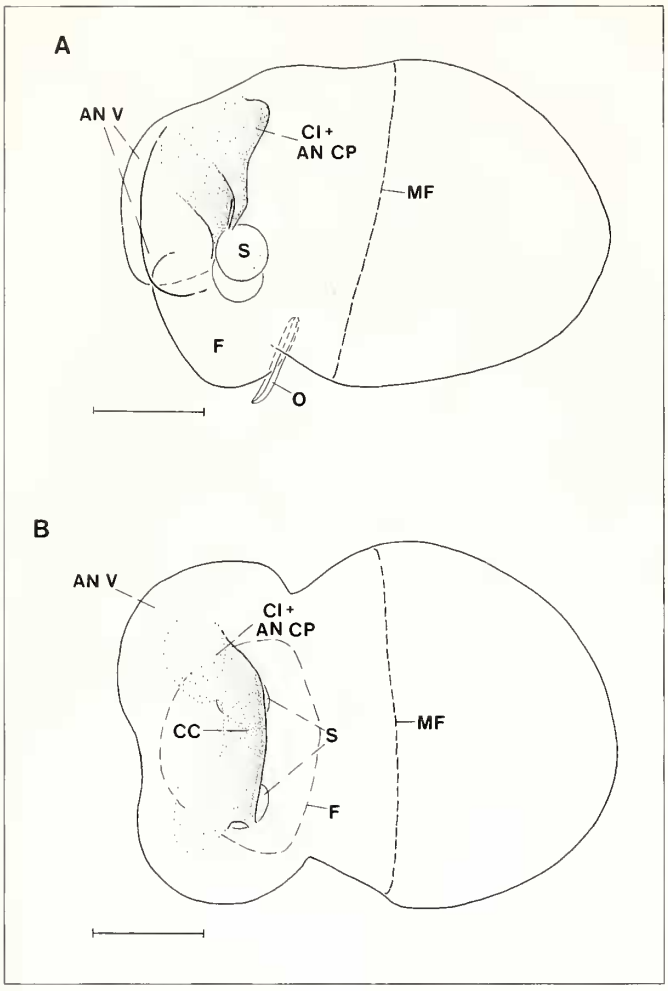

Fig. 2. Reconstruction of the nervous system of a 6-day-old larva of *Ovatella myosotis*.. A. Lateral view from the left side. B. Dorsal view. AN CP + CI = anlage of the cerebropleural ganglia plus cerebral invagination; AN V = anlage of the velum; CC = cerebral commissure; F = foot; MF = mantle fold; O = operculum; S = statocyst. Scale bar = 40 µm.

glion another nerve (Fig. 1, PAN) emerges from the connective connecting suboesophageal and visceral ganglion. The visceral ganglion (Fig. 1, VG) has two large nerves exiting posteriorly. Besides the strong osphradial nerve (Fig. 1, ON), two other nerves (only one visible in Fig. 1) emerge from the supraoesophageal ganglion (Fig. 1, SPG).

(b) Development:

Nervous system:

The paired ganglia of the headfoot and their commissures:

Cerebral and pleural ganglia:

At development day five (day 5 = 5th day after egg deposition), an ectodermal area at the anterior end of the larva, dorsal to the foregut, staines distinctly darker than the surrounding epithelium. It is located directly at the animal pole of the embryo, which is marked by the attached polar bodies. This is the first visible structure of the nervous system and represents the formation zone of the cerebral commissure. It also contributes to the formation of the cerebral and pleural ganglia.

A few hours after the formation of this first anlage, the cerebral commissure can be distinguished at its center. It consists of a row of ectodermal cells, showing dark-staining nervous fibers at their inner base. Subsequently, the ectoderm on both sides of the cerebral commissure thickens. These thickenings lie in the center of each developing velar lobe (Fig. 2). They soon become multicellular by proliferation. Slight invagination troughs are formed in the center of each anlage (Figs. 19, 21), which first appear on development day 7. The structures surrounding the invagination troughs will henceforth be referred to as cerebral invaginations. As predecessors of various sensory and nervous structures in the head area, they play a major role in further development (Fig. 22).

Initially, the cells of the cerebral commissure have a wide

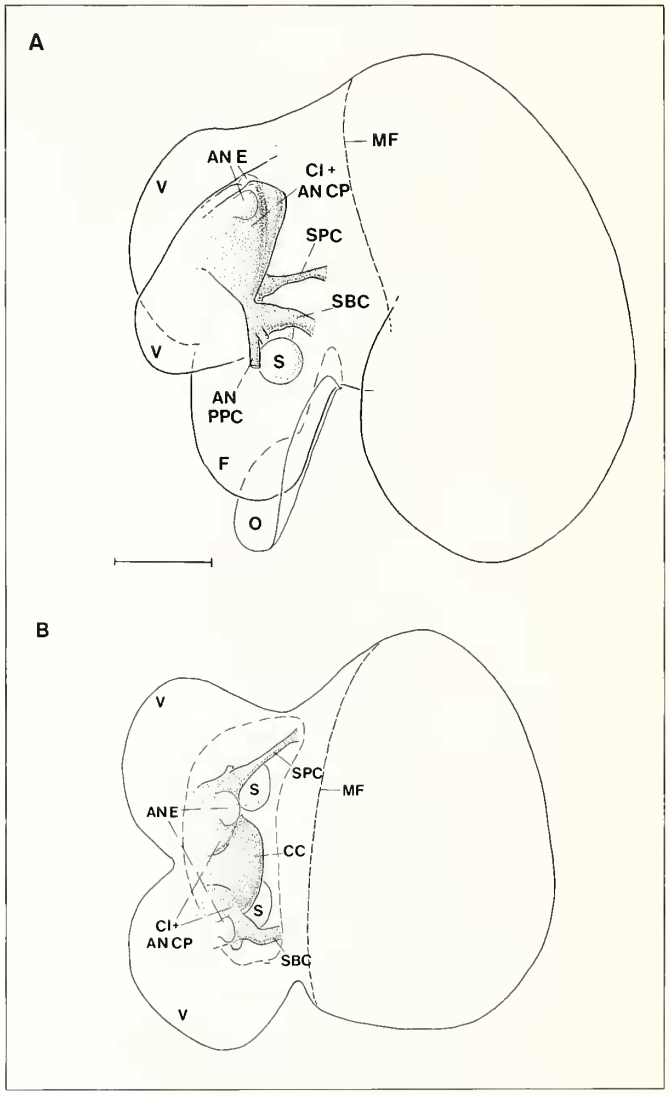

Fig. 3. Reconstruction of the nervous system of an 8-day-old larva. A. Lateral view from the left side. B. Dorsal view. AN CP + CI = anlage of the cerebropleural ganglia plus cerebral invagination; AN E = anlage of the eye; AN PPC = anlage of the pleuropedal connective; CC = cerebral commissure; F = foot; MF = mantle fold; O = operculum; S = statocyst; SBC = suboesophageal connective; SPC = supraoesophageal connective; V = velar lobes. Scale bar = 40 µm.

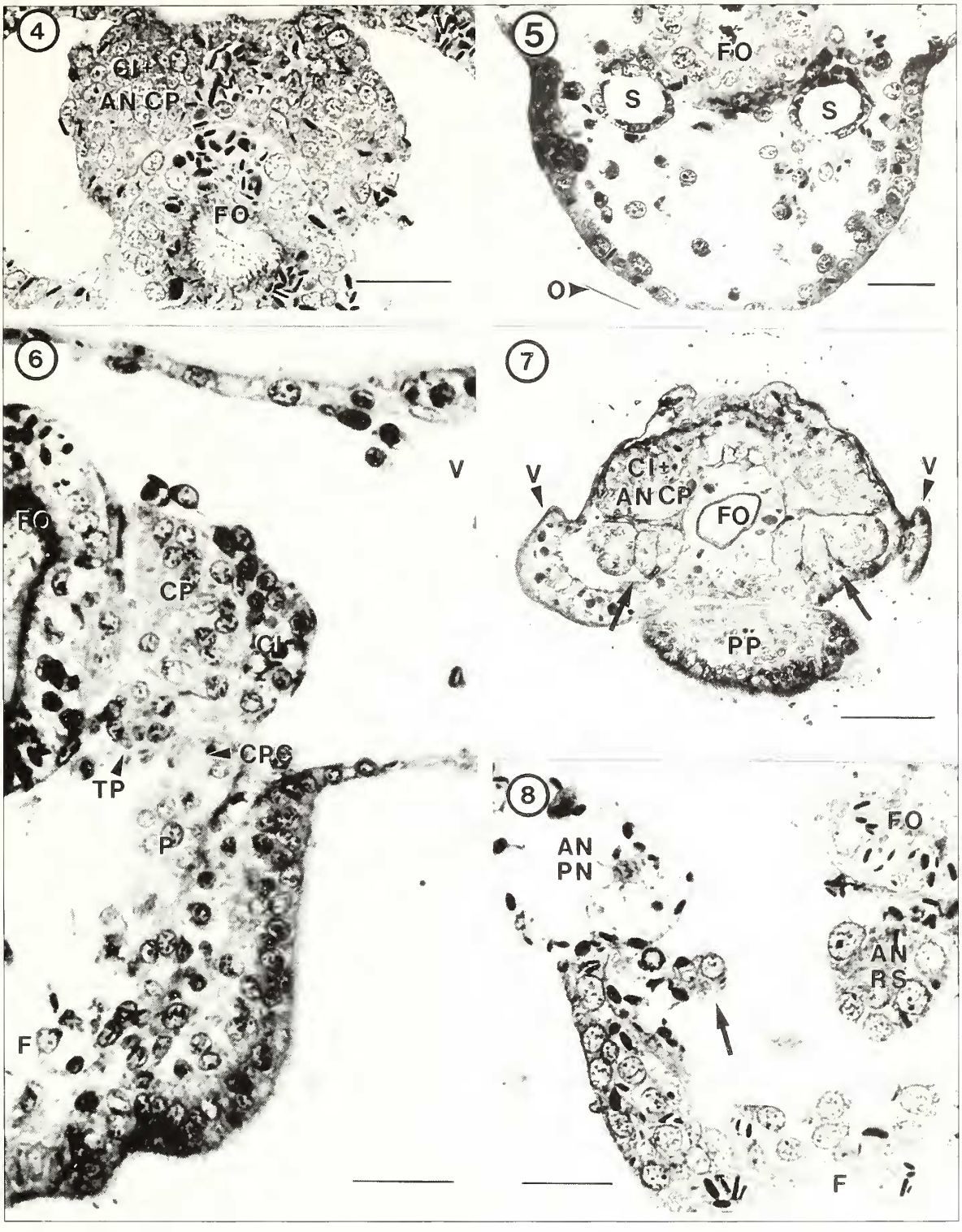

Fig. 4. Cross section through the anterior portion of a 7-day-old larva. AN CP + CI = anlage of the cerebropleural ganglia plus cerebral invagination (only marked on right larval side = left side on figure); FO = foregut; V = velum. Scale bar = 20 µm. Fig. 5. Cross section through the foot of an 8-day-old larva. The lumen contains the cells that form the pedal ganglia. FO = foregut; O = operculum; S = statocysts. Scale bar = 20 µm. Fig. 6. Cross section thtough the anterior nervous system elements of the left side of a 13-day-old larva. CI = cerebral invagination; CP = cerebropleutal ganglion; CPC = cerebropedal connective; F = foot; FO = foregut; P = pedal ganglion; TP = temporary portion of the cerebral ganglion; V = velum. Scale bar = 20 µm. Fig. 7. Cross section through the anterior region of a 12-day-old larva of *Haminoea navicula* showing three pairs of accessory ganglia of the labiotentacular nerve (arrows) (Courtesy of Kurt Schaefer, 1992). AN CP + CI = anlage of the cerebropleural ganglia plus cerebral invagination (only marked on right larval side = left side on figure); FO = foregut; PP = propodium; V = velat lobes. Scale bar = 50 µm. Fig. 8. Cross section through the right side of a 7-day-old larva showing the first anlage of the supraoesophageal ganglion (arrow). AN PN = anlage of the protonephridium; AN RS = anlage of the radula sac; F = foot; FO = foregut. Scale bar = 15 µm.

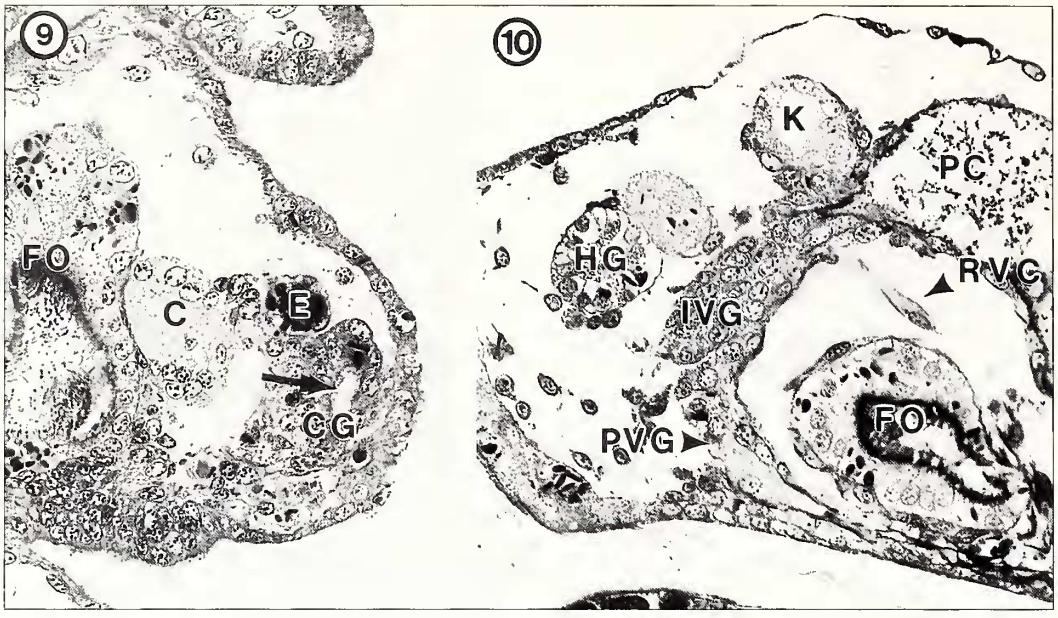

Fig. 9. Cross section through a newly metamorphosed juvenile (15 days) (same specimen as figure 16) in the region of the forming right tentacle. E = eye; FO = foregut; C = cerebral ganglion; CE = cerebral gland; arrow shows the lumen of the forming cerebral gland. Scale bar = 20 µm. Fig. 10. Cross section through a 12-day-old larva showing the region of the forming visceral ganglion. FO = foregut; HG = hindgut; IVG = invaginated portion of the visceral ganglion; K= kidney; PC = pallial cavity; PVG = proliferated portion of the visceral ganglion; RVC = right visceral connective. Scale bar = 20 µm.

cell body and are connected to the ectoderm. The commissure fibers are continuous with fibers which project ventrally. Some of these fibers project towards the statocyst, the others project towards the site of the forming pedal ganglia. As will be shown, the latter nerve branch represents the anlage of the pleuropedal connective (Fig. 3A, AN PPC). A connection with the statocysts is first recognisable by the end of day 5. The inner portion of the cerebral invaginations is in intimate contact with nervous material which is continuous with the cerebral commissure and the ventrally projecting nerve. As will be shown, this nervous material represents the common anlage of the cerebral ganglia (except the procerebrum) and pleural ganglia. This common anlage will henceforth be referred to as the cerebropleural ganglia. The cells for the anlage apparently originate from the posterior portion of the cerebral invaginations, where rapid cell proliferation takes place. This assumed cell migration from the cerebral invagination to the anlage of the cerebropleural ganglia may explain why no boundary is visible between the two structures (Fig. 4).

The cerebral invaginations soon project conspicuously inwards (Fig. 21). Mitotic figures are abundant in sections of these structures. The cerebral invaginations consist of densely packed cells without nervous fibers. The anlagen of the cerebropleural ganglia have fewer cell bodies and consist mainly of nervous fibers (Fig. 14, CP).

On development day 6 the cerebral commissure becomes separated from the epidermis and displaced interiorly. A single, median connection to the epidermis is maintained. This connection is retained throughout the remaining veliger stage. The cerebral commissure contains cell bodies over its entire length throughout the larval period. The nuclei lie on the side of the cells which is closer to the exterior of the animal. No trace of a cephalic sensory organ could be found in the middle of the cerebral commissure and the overlying ectoderm, the site of the usual location of this organ in gastropod larvae.

The cerebropleural ganglia grow until day 9, when they extend ventrally halfway down the oesophagus. They then start to undergo differentiation processes which lead to the formation of the adult elements as well as an additional structure which is only temporary present. The latter, referred to as the temporary portion, separates from the median side of the ganglion anlage on day 11. It is most conspicuous on 13 day, when it is spherical and contains 15-20 nuclei arranged on its outer layer (Fig. 6, TP). It apparently plays a role in the formation of certain nerves (see below). The temporary portion is present until the hatching stage, when it becomes fused with the remaining cerebral ganglion.

Shortly after the differentiation of the temporary portion, the cerebral invagination (= the prospective procerebrum plus cerebral gland) becomes distinctly separated from the rest of the cerebropleural ganglion (Fig. 6, 22C). A membrane forms between the two structures; it may be related to the formation of a cover membrane around the whole ganglion. Although undetectable in earlier developmental stages the separation of the cerebral invagination from the remaining ganglionic anlage may have existed earlier. Both procerebral connectives (first visible on day 13) develop from the junction of the posterior end of the procerebrum-cerebral gland anlage with the remaining ganglion.

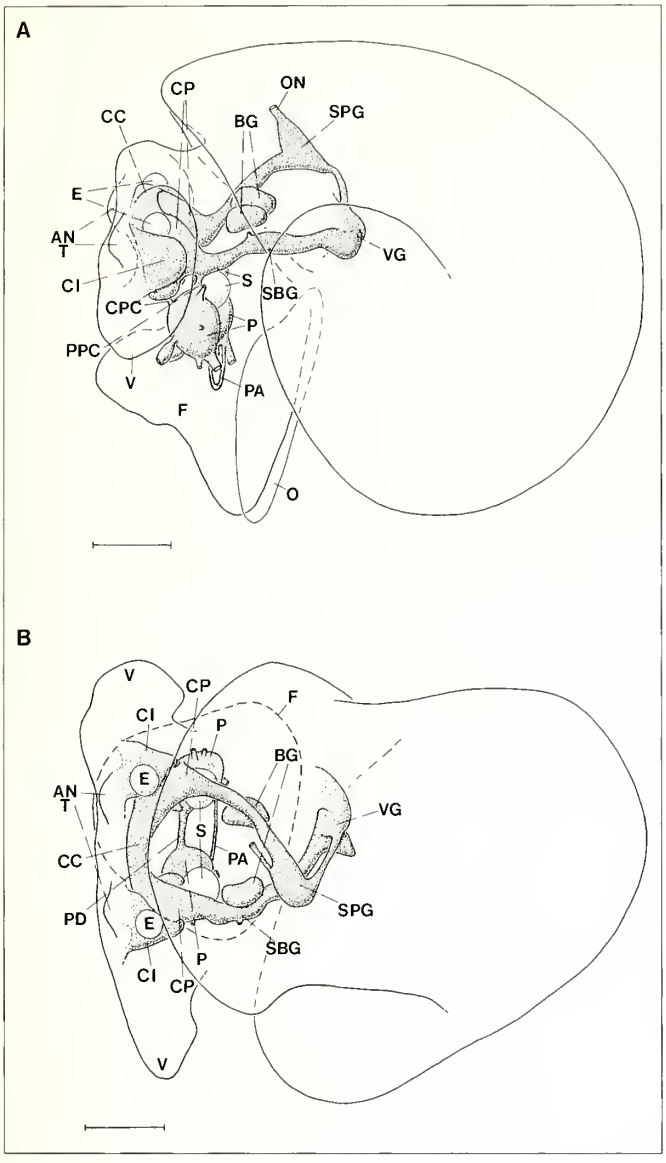

Fig. 11. Reconstruction of the nervous system of a 13-day-old larva. A. Lateral view from the left side. B. Dorsal view. AN T = anlage of the tentacles; BG = buccal ganglion; CC = cerebral commissure; CI = cerebral invagination; CP = cerebropleural ganglion; CPC = cerebropedal connective; E = eye; F = foot; O = operculum; ON = osphradial nerve; P = pedal ganglion; PA = parapedal commissure; PD = pedal commissure; PPC = pleuropedal connective; S = statocyst; SBG = suboesophageal ganglion; SPG = supraoesophageal ganglion; V = velar lobe; VG = visceral ganglion. Scale bar = 50 µm.

At metamorphosis the junction of the procerebrum anlage with the ectoderm and the procerebral invagination troughs narrows considerably; this is caused by the inward movement of the whole ganglionic complex (Fig. 9, 16). This process eventually leads to the condition in which the procerebrum-epidermis connection consists of a narrow tube. Thus, the cerebral tube (the distal portion of the cerebral gland) achieves its typical, elongated, adult shape. At hatching the most distal portion is already tubiform (Fig. 17). This process is illustrated in figure 22.

The pleural ganglia are formed by separation from the larval cerebropleural ganglia. Their initial position corresponds to the site where the pleuropedal connectives emerge from the cerebropleural ganglion. The separation is a slow process beginning on day 11, when the larval cerebropleural ganglion elongates posteriorly. At hatching, separation is completed by a distinct movement of the pleural ganglion in a posterior direction (Fig. 17). At this time the pleural ganglion is still closer to the cerebral than to the pedal ganglion; this represents an epiathroid condition (Fig. 17). The hypoathroid condition, in which the pleural ganglion is closer to the pedal ganglion, is achieved several weeks after hatching. Figure 23 illustrates the formation sequence of the cerebral and pleural ganglia and their connectives to the pedal ganglion.

The first distinguishable elements of the dorsal bodies appear a few weeks after hatching (shell length: 1 mm). These are cells of presumably mesodermal origin attached to the cerebral ganglion near the origin of the cerebral commissure. The dorsal bodies achieve their typical cap-like shape about six weeks (shell length: 2.3 mm) after hatching.

Pedal ganglia:

Immediately prior to the appearance of the pedal ganglia at the end of day 8, sections show a large number of cells in the haemocoel of the foot (Fig. 5). At the same time the laterofrontal epidermis of the foot is thickened and shows cells on its inner base which appear to be detaching from the epidermis. This arrangement of the elements allows the following mode of development for the pedal ganglia to be postulated: Cells detach from the proliferation zones positioned laterofrontally of the foot. These cells individually move to the future position of the pedal ganglion and aggregate there (lateroventral to the statocysts). The pedal ganglia are initially visible as small cell clusters. Subsequently, they temporarily extend to the ectodermal proliferation zones on the side of the foot (day 9-10), at which time there is no visible boundary between ganglia and proliferation zones. On day 11 the ganglia are again detached from the epithelium and become covered with a membrane.

At the end of day 11 the pedal (Fig. 15) and parapedal commissures are formed by the outgrowth of nervous fibers from the inner fibrous portion of the ganglia. The initial outgrowth of the parapedal commissure is posterior to the pedal commissure.

Buccal ganglia:

The formation of the buccal ganglia starts on day 11. They are formed by cells from the dorsal side of the radula sac posterior to its connection with the oesophagus. When first distinguishable, the anlagen are two cell clusters connected anteriorly with the radula sac. The ganglia attain a spherical shape on days 12-13 (figs. 11A,B, 13).

The primordium of the buccal commissure is preformed by cells, although it could not be determined whether these cells originate from the pre-existing buccal ganglia or by proliferation from the radula sac epithelium in between the ganglia. The commissure first appears on day 13 and contains nuclei until metamorphosis (day 15).

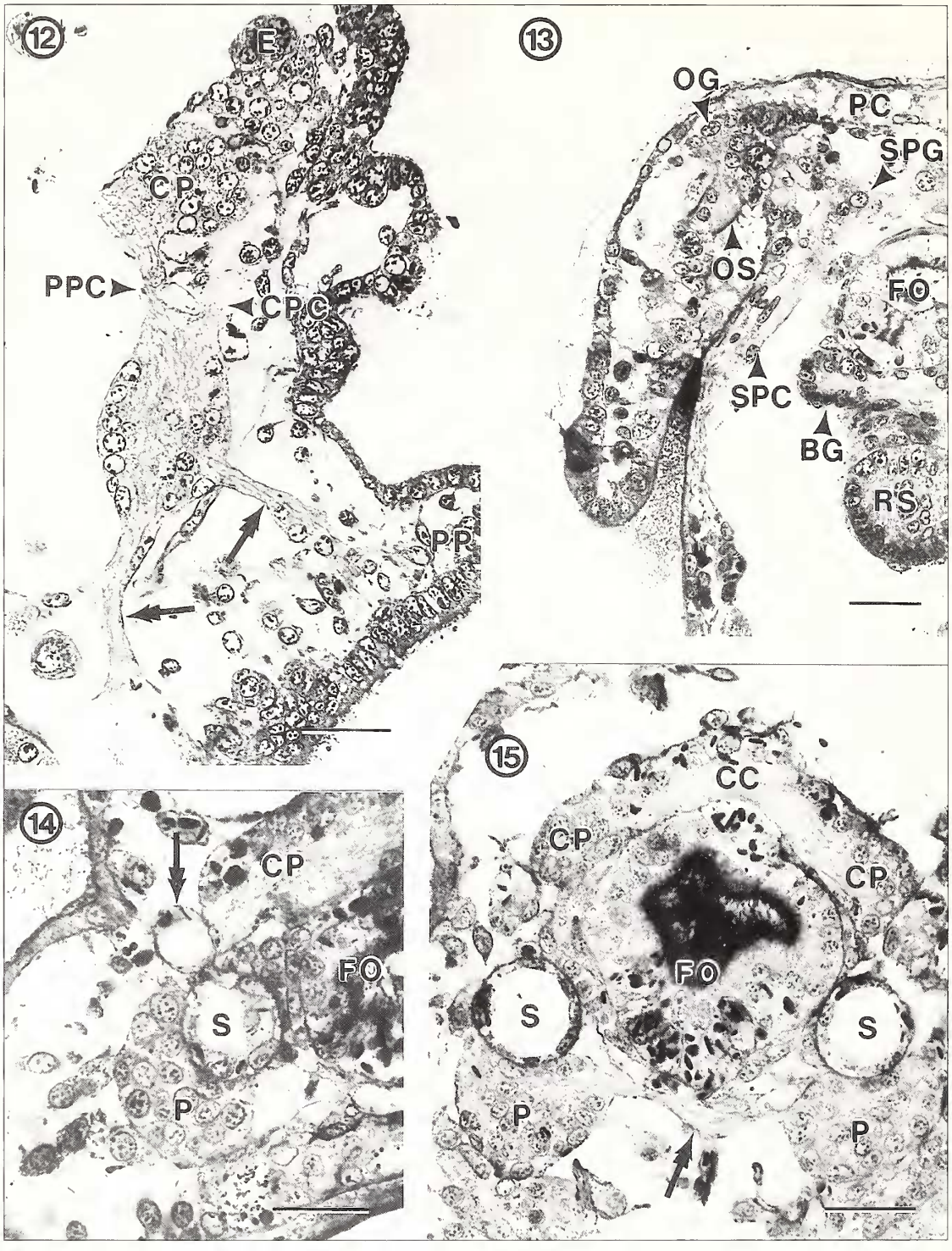

Fig. 12. Parasagittal section through the headfoot of a 13-day-old larva showing two of the pedal nerves (arrows). Note the ganglionic swelling of the anterior pedal nerve in the propodium. CP = cerebropleural ganglion; CPC = cerebropedal connective; E = eye; P = pedal ganglion; PP = propodium; PPC = pleuropedal connective. Scale bar = 20 µm. Fig. 13. Cross section through the right side of a metamorphic competent (day 15) larva. BG = buccal ganglion; OG = osphradial ganglion; OS = osphradium; PC = pallial cavity; RS = radula sac; SPC = supraoesophageal connective, SPG = supraoesophageal ganglion. Scale bar = 20 µm. Fig. 14. Cross section through the right side of a 12-day-old larva showing the static canal of the statocyst (arrow). CP = cerebropleural ganglion; FO = foregut; P = pedal ganglion; S = statocyst. Scale bar = 20 µm. Fig. 15. Cross section through the pedal ganglia of a 12-day-old larva showing the newly formed pedal commissure (arrow). CC = cerebral commissure; CP = cerebropleural ganglion; FO = foregut; P = pedal ganglia; S = statocysts. Scale bar = 20 µm.

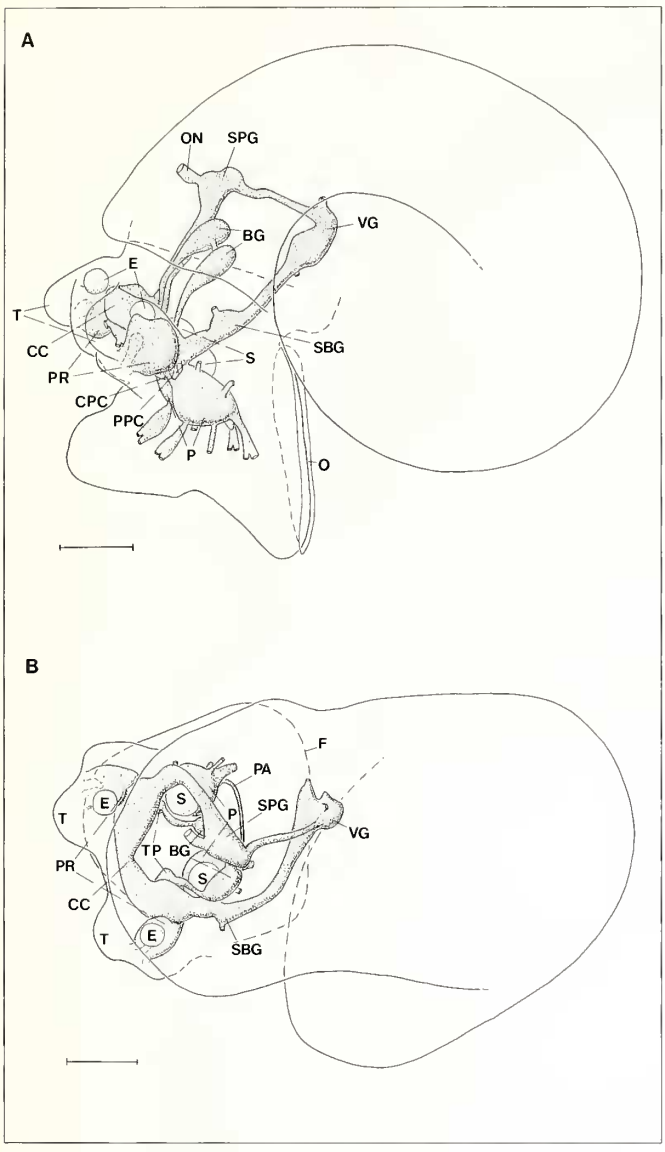

Fig. 16. Reconstruction of the nervous system of a newly metamorphosed juvenile (15 days) A. Lateral view from the left side. B. Dorsal view. BG = buccal ganglion; CC = cerebral commissure; CPC = cerebropedal connective; E = eye; F = foot; O = operculum; ON = osphradial nerve; P = pedal ganglion; PA = parapedal commissure; PPC = pleuropedal connective; PR = procerebrum; S = statocyst; SBG = suboesophageal ganglion; SPG = supraoesophageal ganglion; T = tentacle; TP = temporary portion of the cerebral ganglion; VG = visceral ganglion. Scale bar = 50 µm.

Unpaired ganglia of the visceral portion:

Supraoesophageal ganglion:

The supraoesophageal ganglion is formed through a combination of both invagination and proliferation. The initial primordium appears on day 7 as an aggregation of three to four loosely connected cells directly below the right protonephridium (Fig. 8). These cells presumably originate form an ectodermal proliferation zone slightly dorsal and posterior to the right protonephridium. The primordium is connected with the right cerebropleural ganglion via a connective, which has also formed on day 7. The proliferated portion barely grows until the onset of mantle cavity invagination on day 11. Prior to the mantle cavity invagination, the formation zone of the supraoesophageal ganglion adjoins that of the visceral ganglion. The invagination shifts the supraoesophageal ganglion anlage inside the mantle cavity, where it becomes located in the dorsal epithelium, distinctly to the left of the visceral ganglion. Subsequently, during day 11 the proliferation zone invaginates towards and connects with the already formed portion of the ganglion. Meanwhile the proliferated por-

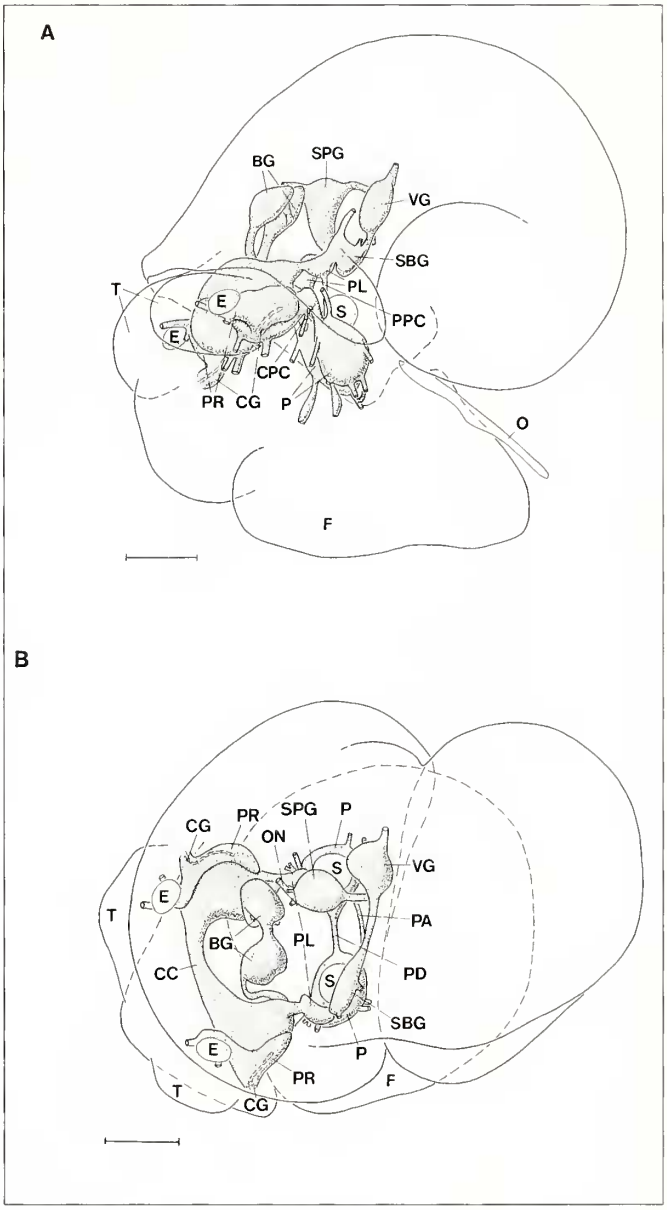

Fig. 17. Reconstruction of the nervous system of a hatching stage juvenile (21 days) A. Lateral view from the left side. B. Dorsal view. BG = buccal ganglion; CC = cerebral commissure; CG = cerebral gland; CPC = cerebropedal connective; E = eye; F = foot; O = operculum; ON = osphradial nerve; P = pedal ganglion; PA = parapedal commissure; PD = pedal commissure; PL = pleural ganglion; PPC = pleuropedal connective; PR = procerebrum; S = statocyst; SBG = suboesophageal ganglion; SPG = supraoesophageal ganglion; T = tentacle; VG = visceral ganglion. Scale bar = 50 µm.

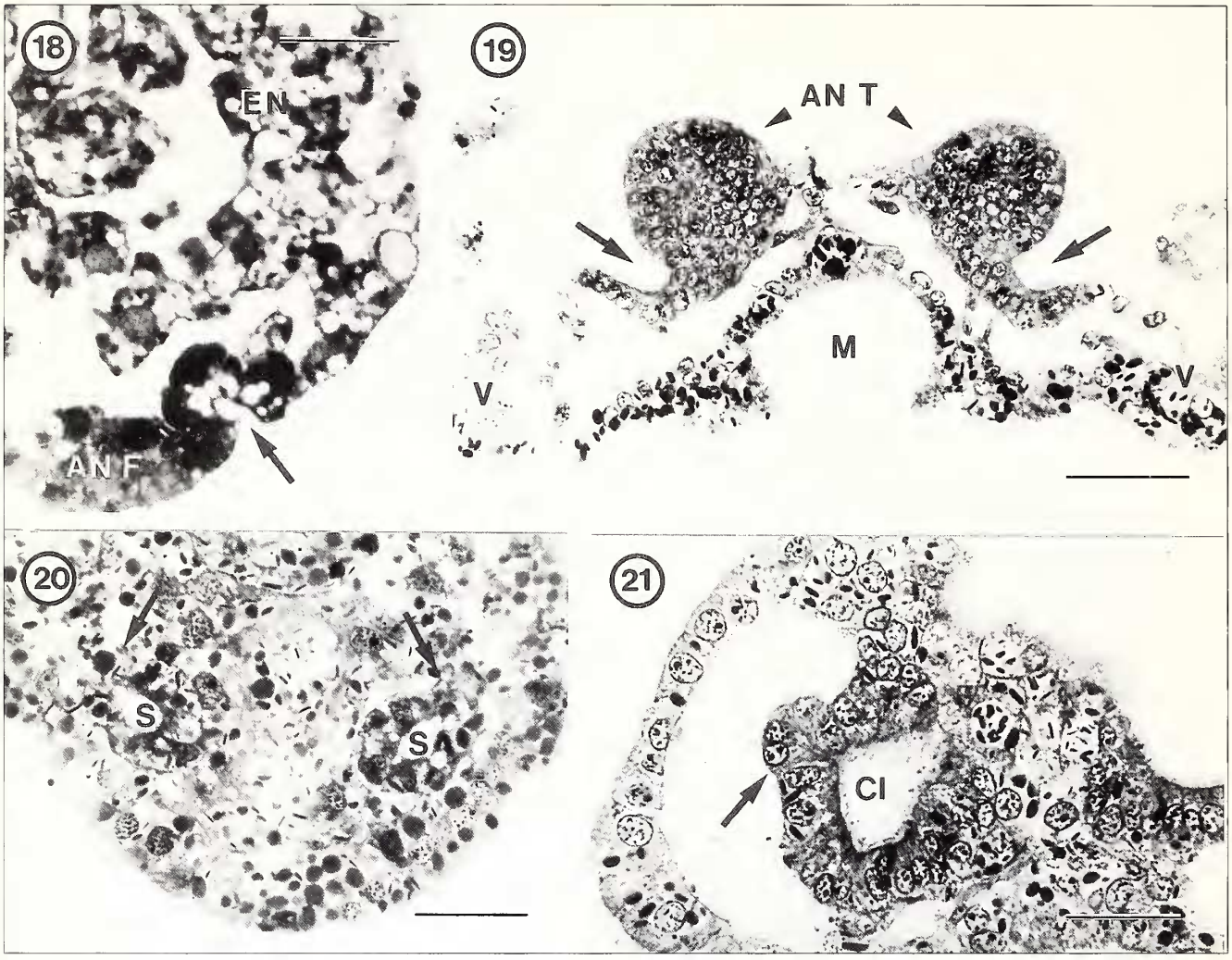

Fig. 18. Cross section through of a 90-hour-old embryo showing the invagination of the left statocyst (arrow). AN F = anlage of the foot; EN = endoderm. Scale bar = 20 µm. Fig. 19. Cross section through the anterior end of a 13-day-old larva showing the relation of the forming tentacles and the opening of the cerebral invaginations (arrows). AN T = anlage of the tentacles; M = mouth opening; V = velar lobes. Scale bar = 30 µm. Fig. 20. Cross section through the foot of a 4-day-old preveliger. Note the dorsolaterally separating portion of the statocysts (arrows). S = statocyst. Scale bar = 20 µm. Fig. 21. Cross section through the right velar lobe and cerebral invagination of an 8-day-old larva showing the invagination of the eye (arrow). CI = cerebral invagination. Scale bar = 20 µm.

tion grows by the attachment of additional cells and becomes compact. Both portions subsequently fuse completely and lose the connection with the ectoderm during day 13.

Visceral ganglion:
The visceral ganglion formation is a combination of the same two processes as in the supraoesophageal ganglion, but in reversed sequence. It starts with an invagination process. The invagination zone is first detectable on day 8 by tracing the fibers of the early-formed left visceral connective. It is positioned in the innermost region of the mantle cavity, which has begun to grow inwards. Thickened epithelial cells form a shallow invagination. Adjacent to this site a mesodermal band projects inwards; it may be confused with elements of the ganglionic anlage. Further invagination of the mantle cavity shifts the anlage of the visceral ganglion posteriorly to a location in the ventral-most epithelium. Subsequently, cells ingress from the invaginated anlage of the ganglion to form a cluster near their site of origin. A narrow junction forms between that cell cluster and the earlier formed epithelial portion of the visceral ganglion anlage, although the cell cluster remains separate. When it first appears, the connective to the supraoesophageal ganglion (right visceral connective) merges with the epithelial, invaginated portion (Fig. 10). On day 12 nervous fibers bridge the junction between the epithelial portion and the inner cell cluster. Figures 10 (IVG, PVG) and 11 (VG) show the two portions of the forming visceral ganglion. Only a small part of the epithelial portion is involved in the subsequent fusion process. The tissue adjacent to the site where the right visceral connective merges with the mantle cavity epithelium fuses with the inner anlage. On day 14 the entire ganglion detaches from the ectoderm.

Suboesophageal ganglion:
The primotdium of the suboesophageal ganglion is first detectable on day 8. It is represented by two cells located ventrally to the left protonephridium at the posterior end of the pleurosuboesophageal connective. The origin of these cells, which subsequently increase in numbet, is unclear. They may stem from the pleural ganglion; no associated ectodetmal formation zone could be detected. When the nervous cord from the left pleural to the viscetal ganglion is completed (day 10), cells attributable to the suboesophageal ganglion are distributed along most of the junction. The nuclei extend from the left cerebropleutal ganglion almost until the visceral ganglion. Until hatching (day 21) there is no distinct ganglion-like thickening visible on that nervous junction (Fig. 11, 16, SBG). At this time, a shift of neurons forms a fibrous connective to the pleural ganglion. Cutrently, the suboesophageal ganglion becomes spherical and grows conspicuously. The formation of the suboesophageal ganglion clearly differs substantially from that of the supraoesophageal and visceral ganglion. The basic difference is the lack of a detectable, associated formation zone. Though one cannot definitely exclude the existence of such a formation zone, the lack of an invagination process is certain. Another major difference is the late differentiation into the typical ganglionic shape, which only occurs after hatching.

Osphradial ganglion:
Cells which proliferate from the earlier fotmed osphtadial epithelium (see below) start forming the osphradial ganglion on day 12 (Fig. 13, OG). At metamorphosis (day 15) it consists of a distinct cluster of cells in close contact with the osphradium. The diffetentiation of a neuropile is first visible on day 18. The ganglion detaches from the epidermis during juvenile development aftet hatching and petsists near its origin throughout life.

Connectives:
Connectives of the headfoot:
The first connectives to be formed are the pleuropedal connectives. They descend from outgtowths projecting ventrally

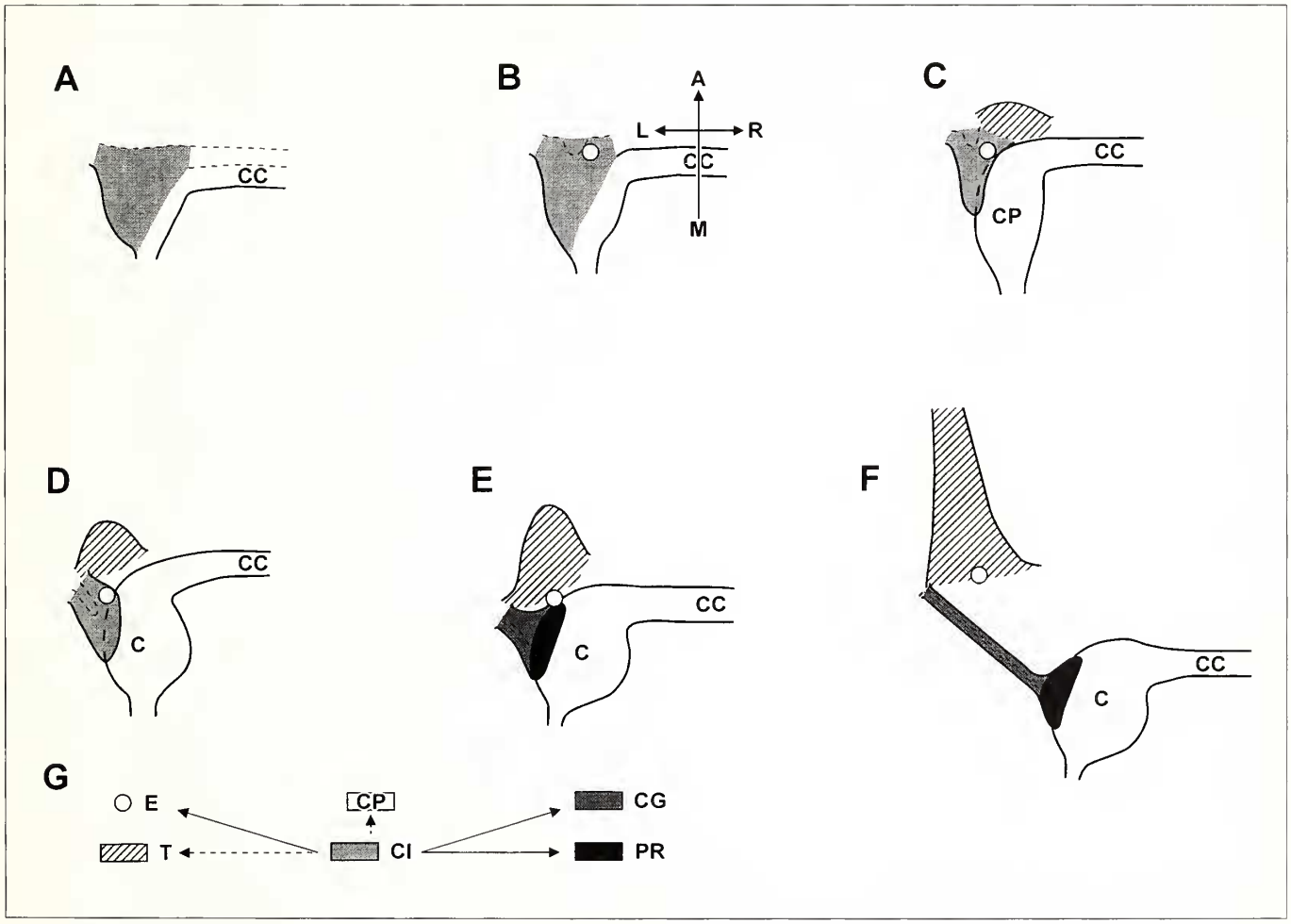

Fig. 22. Diagrammatic sequence of the development of sensory organs and nervous system elements in the head region of *Ovatella myosotis*. A. - F. Dorsal view of the left side; A. Early veliger (day 6) B. Veliger (day 11); C. Late veliger (day 13); D. Postmetamorphic juvenile (day 15) E. Hatching stage; F. Adult; G. Legend for structures derived from the cerebral invagination in A. - F; Stippled arrow if direct derivation is unclear. A = anterior; C = cerebral ganglion; CC = cerebral commissure; CG = cerebral gland; CI = cerebral invagination; CP = cerebropleural ganglion; E = eye; L = left; M = median plane; PR = procerebrum; R = right; T = (anlage of) tentacle.

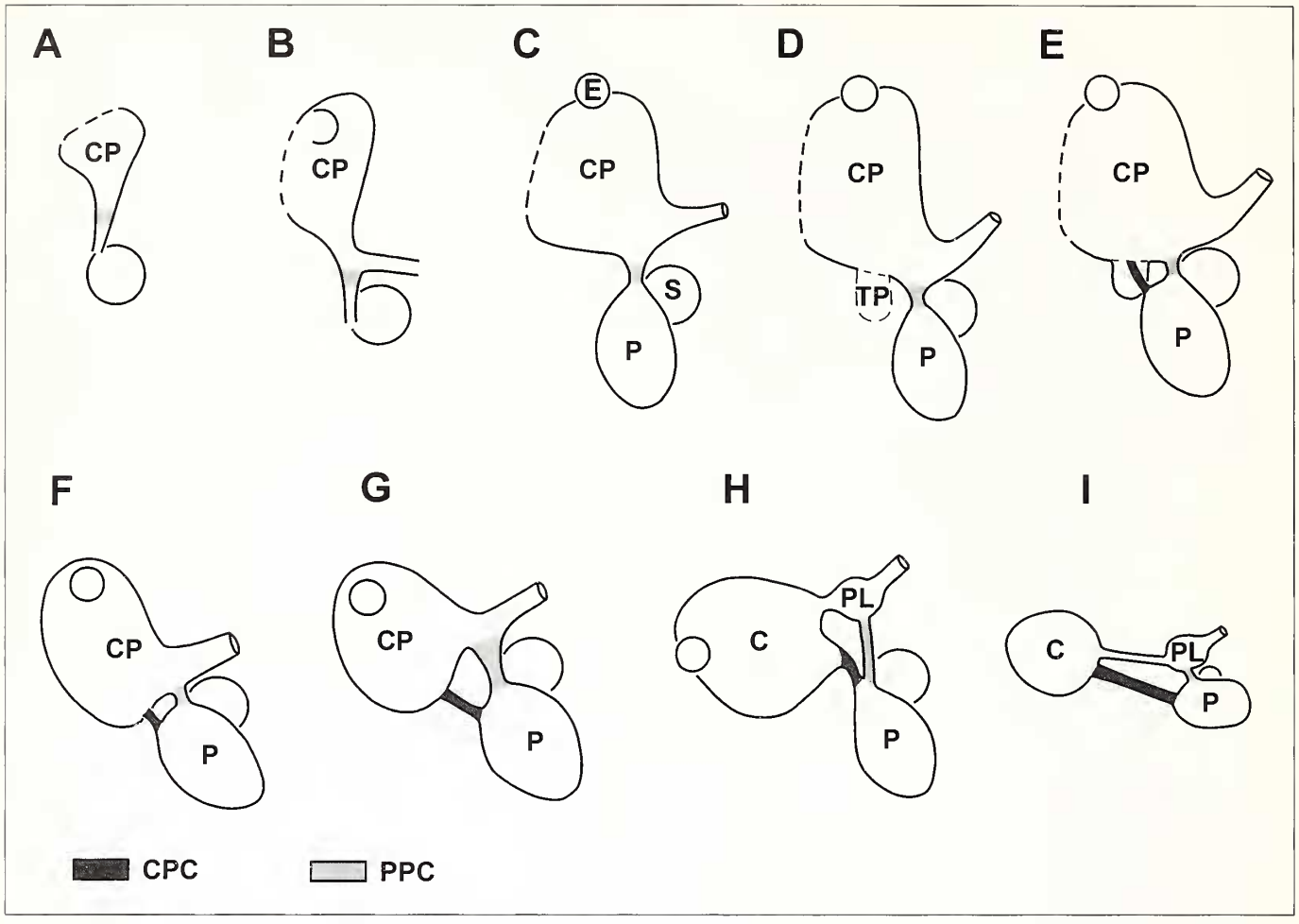

Fig. 23. Diagrammatic sequence of the development of cerebral, pedal and pleural ganglia and the respective connectives of *Ovatella myosotis* (details of cerebral invagination and derived structures not shown). Lateral view from the left side. A. Early veliger (day 6); B. Veliger (day 8); C. Veliger (day 10); D. Veliger (day 11); E. Veliger (day 12); F. Postmetamorphic juvenile (day 15); G. intracapsular juvenile (day 19); H. Hatching stage; I. Adult. C = cerebral ganglion; CP =(anlage of) cerebropleural ganglion; CPC = cerebropedal connective; E = eye (only marked in C.); P = pedal ganglion; PL = pleural ganglion; PPC = pleuropedal connective; S = statocyst (only marked in C.); TP = temporary portion of the cerebral ganglion (only marked in D.). Note that the first-formed connective to the pedal ganglion is the future pleuropedal connective.

from the cerebropleural ganglia. These outgrowths (Fig. 2A) appear shortly after ganglion formation and are initially composed of cells. The two ramifications of the nervous outgrowth extend to the statocysts and to the ectoderm lateral to the statocysts (Fig. 3A). Later (day 10) another ramification, which becomes fibrous in its ventral portion, connects with the forming pedal ganglia. These (paired) ramifications represent the ventral portions of the pleuropedal connectives. During the larval stage, the whole length of the relatively short connectives adjoins the statocysts. They lie anterolateral to the statocysts (Figs. 11A, 16A, 17A). The association of the pleuropedal connective with the statocyst is retained throughout development.

The cerebropedal connectives are first detectable early on day 11 as thin fibrous nerves. They emerge laterally from the cerebral ganglion (Fig. 11A) between the temporary portion and the remaining cerebral ganglion (Fig. 6, CPC). They join the pedal ganglia in a more anterior position than the pleuropedal connectives (Figs. 11A, 12). They may originate as an outgrowth of nervous fibers from the cerebral ganglia towards the pedal ganglia, because in the respective zones, nervous fibers are differentiated later in the pedal than in the cerebral ganglia.

The cerebropleural connectives represent the remainder of the junction between the separating cerebral and pleural ganglia. Shortly before hatching they resemble typical connectives consisting mainly of fibers. The last neurons disappear two days after hatching. Figure 23 provides a semi-schematic outline of the formation mode of the commissures that connect cerebral, pleural and pedal ganglia.

The buccal connectives are first visible prior to metamorphosis. At this time they emerge from the temporary portion of the cerebral ganglion. Figure 16B shows this condition in a postmetamorphic juvenile.

Connectives of the visceral loop:
The supraoesophageal connective connecting the right pleural and supraoesophageal ganglion is formed by a thin outgrowth of cells (day 7). It grows from the pleural portion of the still compact cerebropleural ganglion towards the primordium of the supraoesophageal ganglion (Fig. 3B).

With a slight delay the suboesophageal connective between the left pleural and suboesophageal ganglion is formed in a similar manner. It is first visible on day 8 as a short projection with a terminal thickening, which represents the primordium of the suboesophageal ganglion (Fig. 3B).

When it initially appears on day 10, the right visceral connective is composed solely of fibers and runs adjacent to the epidermis. It passes from the supraoesophageal ganglion to the forming visceral ganglion. Its formation prior to the visceral ganglion indicates a possible origin from outgrowing axons of the supraoesophageal ganglion. Because of the above-mentioned shift of the visceral ganglion it become elongated and is detached from the epithelium during day 11. The nerve then passes to the epidermal site, which is later fused with the proliferated portion of the visceral ganglion (Fig. 11).

The left visceral connective is formed by two elements. The first detectable structures are nervous fibers emerging from the anlage of the visceral ganglion on day 8. They run adjacent to the epidermis and project ventrally. The anterior portion is formed by neurons derived from the suboesophageal ganglion.

The connective later (day 10) becomes fibrous when the neurons aggregate to form the suboesophageal ganglion.

The osphradial connective between the supraoesophageal and osphradial ganglion is initially fairly short (day 12) and of cellular structure. Two days later, after the neurons were dislocated into the osphradial ganglion, it becomes fibrous.

Peripheral nerves (abbreviations refer to the adult in Fig. 1):
The statocyst nerve is not recognisable as a separate nerve until hatching. Like in the adult it is very thin and fibrous and emerges from the cerebral ganglion in the hatching stage. It appears to undergo the following way of development: Beginning on day 5, the statocyst is innervated by branches of the nerve trunk, which also gives rise to the pleuropedal connective (Fig. 3A). The fibers innervating the statocysts presumably split upwards from this trunk before the larval pleural ganglion separates from the cerebropleural ganglion. Ventrally, the statocyst nerve maintains a close contact with the pleuropedal connective (see above).

Most other nerves apparently develop as outgrowing axons from pre-existing ganglia, because they are fibrous when first visible. Their development must be rapid, because halfway extended nerves were never detected.

Two tentacle nerves are present as soon as the cerebral ganglion detaches from the epidermis (after metamorphosis, day 15). They have a common origin from the procerebrum next to the eye. During juvenile development their common base extends so that their bifurcation becomes to lie in some distance to the procerebrum.

Two of the three labial nerves (L1, L2) are present after metamorphosis (Fig. 16A). The third one (L3) appears on day 18. During juvenile development the innervation area of the right labial nerve (L1) becomes the penis complex.

The formation sequence of the ventral pedal nerves proceeds from posterior to anterior. The first-formed (day 11) ventral pedal nerve (VP3) initially emerges at the same point as the parapedal commissure. Soon thereafter, two additional ventral nerves appear (VP1, VP2). The most anterior pedal nerve (VP1) (first visible on day 13) contains a number of neurons, which form a swelling below the pedal ganglion. These neurons are present until hatching and temporarily form an accessory ganglion. This structure and other pedal nerves can be seen in figures 11A, 12, 16A, 17A. The three lateral pedal nerves appear between days 11 and 17. The first one (LP2) extends dorsally. At day 12, a small nerve (LP3) emerges posteriorly. The last nerve (LP1) is the most anterior one (Fig. 17A).

The nerve of the suboesophageal ganglion (SBN) is first detectable after metamorphosis. At hatching another nerve (PAN) becomes visible; it emerges posteriorly from the suboesophageal ganglion (Fig. 17A,B). During post juvenile development the origin of this nerve is shifted posteriorly from the suboesophageal ganglion. The two nerves of the visceral ganglion first appear after metamorphosis (Fig. 16A,B). The osphradial nerve is present shortly after the formation of the supraoesophageal ganglion (day 12). The other supraoesophageal ganglion nerves appear during posthatching juvenile development.

Sensory organs:
Tentacles:
The tentacles arise as exteriorly directed outgrowths of the median lips of the cerebral invaginations (Fig. 19, 22). They are first visible as slight elevations at day 13 (Figs. 11, 19) and become small knobs after metamorphosis (Fig. 16). They elongate to the typical tentacular shape during post-hatching juvenile development.

Statocysts:
The statocysts are formed by ectodermal invaginations (Fig. 18). Starting on day 4, this very rapid process slightly precedes the foot formation. Epidermal contact is lost early on day 5, at which point the anlage is a spherical cluster 16 µm in diameter of five to six cells with a tiny central lumen (diameter: 4 µm). During day 5 the anlage elongates. Subsequently, a small portion partly separates laterodorsally (Fig. 20); this will be termed static canal. At the beginning of day 6 the cells of the anlage start flattening. This process is completed on day 8 and results in the typical cystic shape of the statocyst. Maximum statocyst diameter in the veliger larva is 27 µm. The wall of the static canal attains the same shape (Fig. 14). This structure, which has not yet been reported in ellobiids, persists as a junction between the statocyst and the statocyst nerve in the adult. The ciliation in the statocyst and static canal is first detectable at day 6. The statolith is formed prior to cell flattening. During the veliger stage it grows to about 10 µm. After hatching, the statolith disappears and is replaced by small statoconia (diameter: 2 µm).

Eyes:
Similarly to the statocysts, the eyes are formed by invagination (beginning of day 8). The invagination site is located anterolaterally on the cerebral invagination (Fig. 21). The anlage soon becomes spherical and maintains a lumen, which represents the remainder of the invagination. This lumen enlarges as cells begin to flatten. On day 9, pigment is first deposited on the inner base of the cells. Subsequently, the lens replaces the central lumen. By the end of the veliger phase the lens loses its central position and is located anteriorly inside the eye. The eye formation site becomes incorporated into the procerebrum. One week after hatching the eyes detach from the procerebrum.

Osphradium:
On day 11 a cell with long cilia appears in the epidermis near the opening of the newly formed mantle cavity. It lies close to the giant excretory cell of the right protonephridium and is connected with the anlage of the supraoesophageal ganglion. Subsequently, other densely ciliated cells, are formed. They are columnar, with a basal nucleus. Between day 13 and 16 the outer surface of these cells become slightly protruding. This organ, on the basis of its organisation and innervation, can be identified as the chemoreceptory osphradium (Fig. 13).

The reduction of the osphradium, which is absent in the adults, starts after metamorphosis, simultaneously with the formation of the osphradial ganglion. The cells lose their ciliation and become indistinguishable from the surrounding epithelium. Traces of the osphradial epithelium can be found until development day 18.

DISCUSSION

(a) Adult nervous system of *Ovatella myosotis* and terminology:
Previous descriptions of the gross anatomy of the *Ovatella myosotis* nervous system have been provided by PELSENEER (1894), MEYER (1955) and MARTINS (1996) and differ only slightly from the present investigation. The clear hypoathroid position of the pleural ganglia is recognisable in the figure of Martins only. The reported descending nerves are incomplete and misleading in a some points, as for example Meyer apparently mistook a pedal nerve (herein: LP1) as branch of the statocyst nerve. The terminology used in this paper differs from that of MEYER (1955) and MARTINS (1996) with regard to the parietal ganglion, which is termed the suboesophageal ganglion. MARTINS (1996, Fig. 83) figures a ganglionic swelling with a descending nerve between the "parietal" ganglion and the visceral ganglion which could not be detected in the material investigated in this paper. This structure may be homologous with the base of the pallial nerve (Fig. 1, PAN) lying in the posterior portion of the suboesophageal ganglion. The extreme morphological variability of *O. myosotis* (MARTINS, 1996) may provide an explanation for this difference.

A detailed description of the cerebral ganglion and its nerves was provided by VAN MOL (1967), whose nomenclature is used in this study. He, however, gives incorrect positions for the procerebral connectives (VAN MOL, 1967, Fig. 14), which are described in the median plane of the procerebrum. Also, unlike Van Mol's description, the cerebral commissure emerges medially.

In this study the visceral loop is regarded as the posterior nervous junction between both pleural ganglia and contains the sub-, supraoesophageal and visceral ganglion. This is thought to represent the primitive condition in pulmonates (see discussion). Finally, in contrast with some authors (e.g. BULLOCK, 1965), the pleural ganglia themselves are not regarded as part of the visceral loop.

(b) Justification of the histological techniques for developmental stages:
Light microscopy has two clear advantages over transmission electron microscopy, for the study of organ system development: (a) The investigation of individual specimens requires substantial less effort, allowing a relatively large number of specimens and developmental stages to be examined closely, and the genesis of structures to be followed nearly continuously. (b) Measured three dimensional reconstructions can be prepared therefore enables to follow the development nearly continuously. (c) It is much easier to understand the three dimensional arrangement of components and to prepare detailed measured reconstructions of whole larval nervous systems. Until now, these reconstructions have been lacking in TEM studies of gastropod larvae.

(c) General comparative aspects of gastropod nervous system development:
Cerebral and pleural ganglia:
The first anlagen of the gastropod cerebral or cerebropleural ganglia are thickened ectodermal areas, the cerebral invaginations (also termed cephalic, cerebral or sense plates or "Scheitelplatten"). These paired structures are located in the intravelar area (e. g. *Buccinum*: GIESE, 1978; *Aeolidiella*: TARDY, 1970, 1974; *Melibe*: PAGE, 1992a; *Ovatella*: herein) or in the corresponding position, if no velum is present (e. g. *Pomatias*: CREEK, 1951; *Lymnaea*: REGONDAUD, 1964). This point has found wide agreement among students of gastropod organogenesis. Accounts on subsequent development, however, differ widely. The ganglia are described as being formed by cell proliferation from the cephalic plates or (partly) invagination of the cephalic plates or by a combination of both processes (for a review of the older literature see RAVEN, 1966). The formation of an invagination in the cerebral plates (= cerebral invagination) as in *Ovatella* is common in all gastropod taxa (RAVEN, 1966). Reports vary in regard to what extent the cerebral invaginations contribute to ganglion formation and in the timing of their appearance. The invaginations are regarded as independent structures or as source of material for the ganglia. The latter interpretation is favored in most recent studies (e.g. PAGE, 1992a for *Melibe*; this study for *Ovatella*).

Cerebral ganglion formation in other pulmonates has also

been reported to be very similar to *Ovatella* (*Limax*: HENCHMAN, 1890; *Lymnaea*: REGONDAUD, 1964; RAVEN, 1975). In particular, the formation of the procerebrum and cerebral gland of *Lymnaea* is very similar to that of *Ovatella*.

The temporary median portion of the ganglia in *Ovatella myosotis* (Figs. 6, 16B, TP) resembles the structure Henchman (1890) described for *Limax maximus*. In opisthobranchs, probable homologues were found in *Haminoea navicula* (SCHAEFER, 1992) and *Melibe leonina* (PAGE, 1992a,b). Perhaps one of the two pairs of ganglia in the propodium of dorid nudibranchs is homologous as well, as proposed by PAGE (1993).

Cerebral commissure:
Two completely different ways of cerebral commissure formation have been described in gastropods:
(1) SMITH (1935) for *Patella*, MORITZ (1939) for *Crepidula*, D'ASARO (1966, 1969) for *Thais*, *Bursa* and *Distorsio*, BROWN (1934) for *Philine*, RIEDL (1960) for *Rhodope*, TARDY (1970, 1974) for *Aeolidiella* and SCHAEFER (1992) for *Haminoea* describe it as being preformed by the epithelium between the forming ganglia. This corresponds to my observations in *O. myosotis*.
(2) CONKLIN (1897) for *Crepidula*, DELSMAN (1914) for *Littorina*, CROFTS (1937) for *Haliotis*, ERLANGER (1891a,b, 1892) for *Viviparus* and *Bithynia*, STÖCKMANN-BOSBACH (1989) for *Nucella*, DEMIAN & YOUSIF (1975) for *Marisa*, THOMPSON (1958, 1962) for *Adalaria* and *Tritonia*, SMITH (1967) for *Retusa* and HENCHMAN (1890) for *Limax* describe it as an outgrowth of the pre-existing ganglia.

However, the case of the genus *Crepidula* indicates that some of these observations are erroneous as CONKLIN (1897) or MORITZ (1939) provide completely different descriptions for the same genus. Although CONKLIN (1897) mainly worked on *C. fornica*, he also investigated *C. adunca*, the same species MORITZ (1939) studied. An origin of the cerebral commissure from the epithelium between the ganglia may be generally true in gastropods. This is additionally supported by the presence of the cephalic sensory organ, which may represent a basic feature of gastropod veliger larvae (see below). This organ is innervated by the middle of the cerebral commissure and evidence exists that it is formed prior to the cerebral ganglia (apical organ: SMITH, 1935).

Pleural ganglia:
The opinions on pleural ganglia development of gastropods differ substantially in the literature. These interpretations may be grouped into three categories:
(1) Many authors (e.g. DELSMAN (1914) for *Littorina*, ANDERSEN (1925) for *Viviparus*, CASTEEL (1904) for *Fiona*, RIEDL (1960) for *Rhodope*, SMITH (1967) for *Retusa*, TARDY (1970, 1974) for *Aeolidiella*, KRIEGSTEIN (1977) and JACOB (1984) for *Aplysia*, SCHAEFER (1992) for *Haminoea* and this study for *Ovatella*) found that the pleural ganglia develop from the same anlage as the cerebral ganglion or that both anlagen are intimately connected.
(2) Most of these studies describe a separate origin of the pleural ganglia from an ectodermal proliferation site usually located on the upper side of the foot (e.g. ERLANGER (1892) for *Bithynia*, SMITH (1935) for *Patella*, CROFTS (1937) for *Haliotis*, MORITZ, (1939) for *Crepidula*, CREEK, (1951) for *Pomatias*, D'ASARO (1966, 1969) for *Bursa*, *Distorsio* and *Thais*, HONEGGER (1974) for *Ampullarius*, DEMIAN & YOUSIF (1975) for *Marisa*, GUYOMARC'H-COUSIN (1974) for *Littorina*, THOMPSON (1958) for *Adalaria*, BICKELL & CHIA (1979) for *Doridella*, HENCHMAN (1890) for *Limax* and CUMIN (1972) for *Lymnaea*. In his review on molluscan morphogenesis, RAVEN (1966) assumed that this is the general pattern in gastropods.
(3) GUIART (1899) proposes that pleural ganglia are always formed in connection with the pedal ganglion in gastropods. Recently, PAGE (1992a,b) argues that this is generally true for nudibranchs. She concludes from her study on *Melibe leonina* that cells from separate proliferation zones (placodes) for pleural and pedal ganglia ingress to produce fused pleural and pedal ganglia.

These substantially different descriptions appear to give the impression that the mode of pleural ganglia formation is highly variable and not linked to a specific pattern. However, several reports of a separate origin seem questionable under closer scrutiny. (1) The formation mode of the pedal connectives in *Ovatella myosotis* enables a re-interpretation of certain earlier studies. The fact that the pleuropedal connectives of *Ovatella* are formed prior to the cerebropedal connectives and remain larger than the latter throughout the larval period (Fig. 23) is surprising. It suggests that some authors (e.g. HENCHMAN, 1890; CONKLIN, 1897; KRIEGSTEIN, 1977 (see below); PAGE, 1992a,b (see below)) apparently mistook the pleuropedal connective as the cerebropedal connective. The incorrect interpretation of connectives leads to a false determination of the respective ganglia. (2) Descriptions reporting a separate ectodermal proliferation site for the pleural ganglion are vague and provide no explaining figures. Moreover, some workers admit some doubts on this matter (HENCHMAN, 1890; GUYOMARC'H-COUSIN, 1974). (3) It is generally problematic to identify the path of unlabelled migrating cells, regardless of the histological sectioning technique applied. Attributing an ectodermal proliferation zone (placode) to a forming ganglion becomes more hypothetical the more distantly both structures are located. However, such accounts are the strongest or only evidence for different development, rather than a common origin, of the pleural and cerebral ganglia.

Although data on pleural ganglia formation in gastropods are incomplete and contradictory, they allow speculation on the evolutionary trend of their formation. According to CROFTS' description (1937) of the developing nervous system of *Haliotis*, the pleural ganglia lie next to the pedal ganglia when first visible. The long distance between pleural and cerebral ganglia precludes the formation of the former from a common anlage with the cerebral ganglion in this archaeogastropod. Rather, the pleural ganglia appear to be formed in close association with the pedal ganglia, as observed in another archaeogastropod, *Theodoxus prevostianus* (pers. obs.). This suggests a correlation between the mode of formation and later position of the pleural ganglia. In archaeogastropods the pleural ganglia lie near the pedal ganglia (hypoathroid condition), whereas in other gas-

tropods they (primarily) lie near the cerebral ganglia (epiathroid condition) (a secondary hypoathroid condition as in ellobiids often occurs) (SALVINI-PLAWEN & HASZPRUNAR, 1987; HASZPRUNAR, 1988, 1993). Does the development of the pleural ganglion reflect this condition in that it is formed in association with the pedal ganglion in primitive gastropods and in association with the cerebral ganglion in evolved gastropods? If so, one may generally expect the pleural ganglia to be formed at least in close association with the cerebral ganglia in gastropods with an epiathroid organisation. Most of the data for opisthobranchs and pulmonates correspond to that model. Further observations, in particular on archaeogastropods and caenogastropods, will be needed for a definitive answer. For a discussion of this topic see also HASZPRUNAR (1993).

Pedal commissure:
The literature contains a variety of different accounts of the formation of the gastropod pedal commissures. The outgrowth of nervous fibers from the pedal ganglia was observed in *Littorina* (GUYOMARC'H-COUSIN, 1974) and was similar in *Ovatella*. In *Limax* (HENCHMAN, 1890) and *Haliotis* (CROFTS, 1937) cell outgrowth was observed. In some caenogastropods (MORITZ, 1939; D'ASARO, 1969) the primordium consists of cells closely associated with the overlying ectoderm. These substantially different modes of formation might be explained by the different proportions and developmental sequences of the pedal elements.

Visceral loop:
Previous studies on gastropod visceral loop formation concentrated on the shifting and fusion of complete ganglia (REGONDAUD, 1964; KRIEGSTEIN, 1977). Data on the actual formation of individual elements are scarce.

The information available on the location of the ectodermal placodes of the gastropod visceral loop ganglia differs considerably. In most prosobranchs the placodes are reported to lie near the opening of, or inside, the mantle cavity (SMITH, 1935 for *Patella*; CROFTS, 1937 for *Haliotis*; CREEK, 1951 for *Pomatias*; D'ASARO, 1969 for *Distorsio* and *Bursa*). This probably represents the primitive condition, as in pulmonates (this study for *Ovatella*) and opisthobranchs (PAGE, 1992a for *Melibe*). The reported position of the primordia outside the mantle cavity as in ampullariids (Prosobranchia) (e. g. DEMIAN & YOUSIF, 1975) may be explained by the derived developmental mode of these gastropods (direct development and delayed mantle cavity formation).

The development and organisation of the larval visceral loop of *Melibe* (PAGE, 1992a) resembles that of *Ovatella* in many respects, although Page's interpretations of the "left pallial placode" differs. This structure is closely associated with the "visceral placode". The "left pallial placode" and the "visceral placode" resemble the two portions of the visceral ganglion anlage of *Ovatella*. Page's interpretation may be rooted in her preference for the hypothesis of individual anlagen for all ganglia (p. 359). The above reports of a common anlage of the cerebral and pleural ganglion clearly prove this hypothesis to be incorrect. The account of the late appearance of the "visceral placode" (6 days after shell loss) in *Melibe* (PAGE, 1992a, p. 356) must be erroneous, because elsewhere this structure is figured in an early veliger stage (PAGE, 1992a, fig. 3B).

The developmental mode of the suboesophageal ganglion of *O. myosotis* is remarkable because all previous studies report an individual ectodermal proliferation zone for that ganglion. If the "left pallial placode" of *Melibe* is regarded as part of the visceral ganglion anlage, then the suboesophageal ganglion may be formed in a similar manner as in *Ovatella*.

The newly formed connectives of the gastropod visceral loop have often been reported as bands of cells. They were described either as outgrowths from the ganglia (DELSMAN, 1914; DEMIAN & YOUSIF, 1975) or as formed by the ectoderm located between the forming ganglia (RIEDL, 1960; D'ASARO, 1966, 1969). More recent studies (PAGE, 1992a,b, 1993; this study) found newly formed connectives that consist solely of fibers. This might be due to the much higher resolution of the histological techniques used in this study, suggesting that fibrous connectives may have previously been overlooked.

(d) Comments on previous studies on gastropod nervous system genesis:
KRIEGSTEIN (1977) described the developmental sequence and fusion of ganglia in *Aplysia californica* in detail without, however, particularly focusing on the origin of the pleural ganglia. An examination of his figures (Fig. 1) suggests that what he designated as the "cerebral ganglion" may be the cerebropleural ganglion, and that the first connection between the "cerebral" and pedal ganglion (stage "2") is the true pleuropedal connective. Three indications support this conclusion: (1) JACOB (1984) described a common anlage for cerebral and pleural ganglia in the same species. (2) The angle of the "cerebropedal connective" and its position directly in front of the statocyst in stage "2" is identical to that of the pleuropedal connective in the subsequent stages. (3) A comparison with the corresponding structures of *Ovatella myosotis* (Fig. 23) reveals a remarkable similarity except for the determination of the "cerebral" ganglion and the connective to the pedal ganglion. Kriegstein's interpretation might be due to the fact that he did not examine more stages of the crucial developmental phase between his stages "2" and "3": The detailed examination of exactly this phase led to the different results in *Ovatella*.

JACOB (1984) was the only investigator to use ectodermal cell labeling. Although the study provides decisive support for the widely accepted hypothesis of the ectodermal origin of neurons, the methods may be inappropriate to relate certain proliferation zones to individual forming ganglia. Because Jacob did not specifically label certain ectodermal areas, morphological criteria had to be used to trace individual cells or cell groups. Based on her interpretations, however, this morphological analysis appears to be of limited reliability. For example the structure she designated as "heart" (Figs. 2b, 4b, 5b, 6a; H) is clearly not a heart at all. Furthermore, the presence of cells with internal ciliary bundles (Fig. 8a) is interpreted as indicating ganglia formation from ciliated ectodermal cells. However, only particular epidermal areas like the velar cells or the anterior part

of the foot are ciliated in gastropod veligers: ganglia formation zones are predominantly non-ciliated. The figures in Jacob's study (figs. 3, 4a, 5a) suggest that she mistook velar cells as proliferation zones. An alternative interpretation of the internally ciliated cells in the larval nervous system of *Aplysia* is given by SCHAEFER (1992): they probably represent cells of the cephalic sensory organ of gastropod larvae (see BONAR, 1978; CHIA & KOSS, 1984; UTHE, 1991, 1995). As argued above, the presence of additional lateral proliferation zones for the "cerebral ganglia" (i.e. the cerebral ganglia plus commissure) remains questionable.

The most detailed account on nervous system genesis of gastropods is given by PAGE (1992a,b) for *Melibe leonina*. Unfortunately, interpretation errors led to the striking conclusion that the pleural ganglia are fused with the pedal ganglia throughout development. This conclusion was immediately questioned (HASZPRUNAR, 1993; GOSLINER, 1994; CARROLL & KEMPF, 1994), and in a subsequent paper the author (PAGE, 1993) admitted that her main hypothesis was incorrect. Since details were not provided, it seems necessary to re-examine the model in detail to point out its problems.

In the examined posthatching larval stages, two pairs of connectives were found, projecting ventrally from the "cerebral ganglia" (Fig. 24D). The first-formed pair, which is already present at hatching, was interpreted as the "cerebropedal connectives" and the other one as the "cerebropleural connectives". These interpretations can be disproved by comparative anatomical as well as by embryological evidence. (1) PAGE (1992b) identified the "cerebropedal connectives" because they "...can be identified in all larval and post-larval stages by their association with the statocyst nerves...". This incorrect premise appears to be responsible for the later confusions. In fact, statocysts are typically located adjacent to the pleuropedal connective in gastropods. In adults, the statocysts typically lie partially embedded in the pedal ganglion, posteriorly, adjacent to the junction of the pleuropedal connective and the pedal ganglion (e.g. LEMCHE, 1956, figs. 354, 356; HERSCHLER & DAVIS, 1980, fig. 6; HASZPRUNAR, 1985b, fig. 3, 1985c, fig. 3A). In adult gastropods the pleuropedal connective is therefore much more closely associated with the statocyst than the cerebropedal connective is.. Although the statocyst nerves always emerge from the cerebral ganglion, the ventral portions of these nerves and those of the pleuropedal connectives are in intimate contact with each other, as in *Ovatella*. The nervous system of *Melibe* (PAGE, 1992b, Fig. 1) shows this typical configuration when interpreted in this manner. (2) Features of the development and organisation of anterior nervous system elements of *Ovatella myosotis* show remarkable similarities with those of *Melibe leonina*: The first formed pair of ventral connectives from the cerebral ganglia contains fibers of the statocyst nerve, is thicker and lies posteriorly to the other pair. Because of these similarities, the homology of the respective connectives seems highly likely. Therefore the designations of the connectives to the pedal ganglion of *Melibe* should be changed as follows: The "cerebropedal connective" of Page represents the pleuropedal connective; the "cerebropleural connective" represents the cerebropedal connective.

Page interpreted the portion of the pedal ganglion which forms the connection to the cerebropedal ("cerebropleural") connective as "pleural ganglion" (Fig. 24D). Based on the revised interpretation, the only possible position for the pleural ganglia is where the pleuropedal connectives merge with the dorsal nervous system portion, the cerebropleural ganglion.

Page herself acknowledged that the position of the pleural ganglion is the most controversial part of her analysis. Her interpretation of a portion of the pedal ganglia as the "pleural ganglia" was mainly based on the positions of ectodermal placodes. She assumed that ingression zones (placodes) on the side of the foot of *Melibe* are homologous with prosobranch and pulmonate ingression zones, which reportedly provide cells for the pleural ganglia. The shortcomings of this deduction are: (1) the limited reliability of previous data for a separate ingression site for pleural neurons (see above); (2) the failure to explain how to trace exactly the ingression path of neurons from distantly located placodes in *Melibe*. Therefore the pleural ganglia of *Melibe* - as in any nudibranch - must be regarded as located in between the cerebral and visceral loop ganglia throughout development.

In another study PAGE (1993) used the developmental sequence of nudibranch nervous system elements for far-reaching considerations on molluscan origin and phylogeny. She applied the neutral terms anterior and posterior pedal connectives for the two pairs of connectives that emerge from the pedal ganglion (Fig. 24E) and stated that the posterior connectives emerge from the visceral loop. As explained above, these connectives obviously represent the cerebropedal (anterior) and pleuropedal (posterior) connectives. The sites of their origin from the dorsal portion of the nervous system mark the position of the respective ganglia, which are usually fused in nudibranchs. Thus, the pleural ganglia are located where the posterior (pleuropedal) connectives emerge. Page neglected to mention the pleural ganglia and left the question of their position open, stating that the posterior connectives emerge from the visceral loop. While this at first seems to contradict the designation of the posterior connective as the pleuropedal connective, the problem may simply involve terminological discrepancies because elsewhere, PAGE (1992a, p. 354), following BULLOCK (1965), regarded the pleural ganglia as part of the visceral loop. Accordingly, the posterior connectives would be construed as emerging from the pleural ganglia. Interpreting the pleural ganglia as part of the visceral loop is unusual because most workers regard the visceral loop as emerging from the pleural ganglia (e.g. FRETTER & GRAHAM, 1962; FRANC, 1968; SCHMECKEL, 1985; HASZPRUNAR, 1985a, 1988; SALVINI-PLAWEN, 1991; herein).

PAGE (1993) also questioned conventional models of molluscan and gastropodan tetraneury. This is based on the interpretation that the pedal cords stem from the visceral loop and not from the circumoesophageal nerve ring. However, if the pleural ganglia are regarded as elements of the circumoesophageal nerve ring, no contradiction to the conventional model of gastropodan tetraneury is recognisable: the first-formed structure is the anterior nerve ring with three pairs of ganglia (which may be partly fused); the visceral loop originates from the pleural ganglia, and the pedal nerves from the pedal ganglia. The late appearance of

the anterior (cerebropedal) connectives fits well into that model as they may represent an additional structure to the primary nerve ring. PAGE's (1993) model also identified pedal cords in nudibranch larvae, a surprising claim because advanced gastropods such as opisthobranchs usually do not have pedal cords like primitive gastropods and other molluscs. In the pulmonate *O. myosotis*, for instance, neither larval nor adult anatomy allow one of the nerves to be homologized with the archaeogastropod pedal cords. The lack of a convincing justification for the homology of the first-formed pedal nerves of *Melibe* with archaeogastropod pedal cords makes this proposal doubtful.

CHIA & KOSS (1989) and PAGE (1993) detected two pairs of ganglia in the propodium of late stage dorid nudibranch larvae. PAGE (1993) homologized them with structures of the monoplacophoran nervous system. She viewed the "labial ganglia", which may be homologous to the temporary portion of the larval cerebral ganglion in *Ovatella* (see above), as a homologue of the monoplacophoran labial ganglia plus commissure. Although this is an intriguing idea, it requires additional supportive data from the nervous system genesis of primitive gastropods to be confirmed. The homology of the other ("subradular") pair of ganglia with the subradular ganglion of monoplacophorans (PAGE, 1993) is totally unfounded, and the supposedly homologous structure is not even present in monoplacophorans. New detailed investigations on the nervous system in Monoplacophora (HASZPRUNAR & SCHAEFER, 1996a,b; SCHAEFER & HASZPRUNAR, 1996) revealed that they lack a subradular commissure with a true ganglion. I suggest that the two latter structures are a special feature of dorid nudibranchs, representing one of the numerous pairs of accessory ganglia in the head region of adult dorids (WOLTER, 1967) whose formation was accelerated. This phenomenon of adultation (JÄGERSTEN, 1972; GOULD, 1977) - the shifting of adult characters into larvae - is quite common in nudibranchs as external adult features, which are absent in typical opisthobranch larvae, have been reported for a number of veligers. These include rhinophores in *Rostanga* (CHIA & KOSS, 1982); cerrata anlagen in *Melibe* (BICKELL & KEMPF, 1983); shell loss, rhinophores and cerrata anlagen in *Tergipes* (DE NORDMANN, 1846) and *Aegires* (THIRIOT-QUIÉVREUX, 1977). A comparable case of accelerated gangliogenesis in opisthobranchs are the ganglia lying ventrally, adjacent to the cerebropleural ganglia of the *Haminoea* veliger (Fig. 7). Subsequent development reveals these ganglia to be accelerated (= previously formed) ganglia of the labiotentacular nerve (Schaefer, 1992). The homology of the latter with the ganglia in the propodium of dorid nudibranchs seems unlikely due to their distinctly different location ventral to the oesophagus.

KEMPF *et al.* (1987) and CARROLL & KEMPF (1994) provided another interpretation for the location of the pleural ganglion in the larval nudibranch nervous system. They located the pleural ganglia of *Tritonia diomedea* and *Berghia verrucicornis* (Fig. 24C) in the visceral loop, far from the cerebral ganglia. As already suggested by PAGE (1992b) for *Tritonia diomedea*, these ganglia should be interpreted as visceral loop ganglia because of (1) their equal distance to the "cerebral" and the visceral ganglion, (2) their unequal position in the horizontal plane, as is apparent for *Tritonia* and (3) the lack of connectives to the pedal ganglia. What contributed to the interpretation of KEMPF *et al.*. (1987) and CARROLL & KEMPF (1994)? In aeolidiid nudibranchs the connectives from the larval cerebropleural ganglion to the pedal ganglion are extremely close to each other. In the larval *Aeolidiella soemmeringi* (for taxonomy see SCHMECKEL, 1985) the cerebropedal and pleuropedal connectives are separated by a small gap only (TARDY, 1970, fig. 9A) (Fig. 24A). The same arrangement is present in the aeolidiid *Phestilla sibogae* throughout development (pers. obs.). It therefore seems likely that the connection to the pedal ganglion in the aeolidiid *Berghia verrucicornis* in fact represents both the cerebropedal and pleuropedal connective, which CARROLL & KEMPF (1994) were unable to resolve separately. The thickness of the respective structure in the tritoniid *Tritonia diomedea* (KEMPF *et al.*, 1987, fig. 2D) indicates that the same holds true for this species. As a consequence the designation of *Tritonia* and *Berghia* larval nervous system elements should be altered as follows: the "cerebral ganglia" are the cerebropleural ganglia; the "pleural ganglia" are visceral loop ganglia which may represent the sub- and supraoesophageal ganglia.

Figure 24 gives a comprehensive illustration of descriptions of the anterior nervous system components of late stage euthyneuran larvae, pointing out the varying interpretations of elements. It seems important to note that the designation is unmistakable in *Aplysia californica* (Fig. 24B) and *Ovatella myosotis* (Fig. 24F). Only in these species are the pleural ganglia separate in the adult, allowing clear designation of the surrounding elements. Interpretation of larval elements was done by tracing the fate of the elements in question until the adult. I suggest that the comparison of larval stages of other euthyneurans with these species would prove very helpful in designating the elements in question and lead to very similar results, as is the case in *Aeolidiella* (Fig. 24A) and *Haminaea* (Fig. 24F).

It has been shown here that the pleural ganglia lie between the cerebral ganglia and the viscera loop. This general position is supported by descriptions of separate pleural ganglia in some nudibranchs (MAC FARLAND & O'DONOGHUE, 1929; WÄGELE, 1989). Like in other gastropod groups, this ganglion is clearly separated from the cerebral ganglion but connected with this by the cerebropleural connective, and with the pedal ganglion by the pleuropedal connectives. It remains, however, unclear if these ganglia contain portions of visceral loop ganglia.

(e) Sensory organs:

Gastropod eyes and statocysts typically originate from ectodermal invaginations (e. g. *Marisa*: DEMIAN & YOUSIF, 1975; *Ovatella*: herein). The statocyst anlage is generally positioned on the side of the foot, whereas the eye anlage is near the cerebral ganglion.

The static canal of the opisthobranch larva *Rostanga pulchra* (CHIA *et al.*, 1981) is very similar to that of *O. myosotis* in position, organisation and relation to the statocyst nerve, suggesting a homology of the structures. In *O. myosotis*, however, it does not represent the remnant of the statocyst invagination as it was presumed by CHIA et al. (1981) for *R. pulchra*. It is pre-

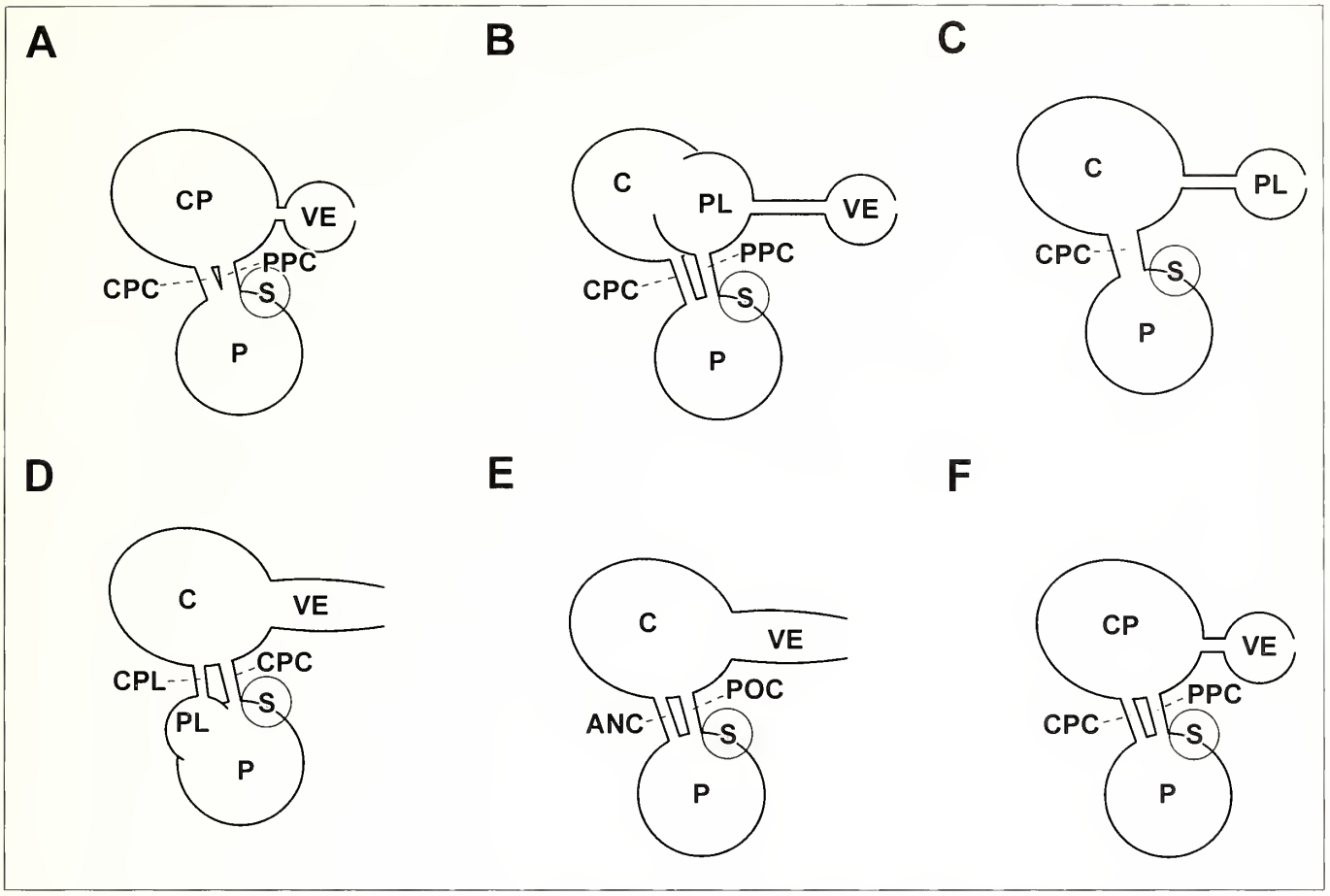

Fig. 24. Diagrammic illustration of descriptions of anterior nervous system elements of late stage euthyneuran larvae. Lateral view from the left. A. *Aeolidiella soemmeringi* (TARDY, 1970, 1974); B. *Aplysia californica* (KRIEGSTEIN, 1977); C. *Berghia verrucicornis* (CARROLL & KEMPF, 1994); D. *Melibe leonina* (PAGE, 1992a,b); E. *Rostanga pulchra* (PAGE, 1993); F. *Haminaea navicula* (SCHAEFER, 1992), *Ovatella myosotis* (herein); ANC = anterior connective; C = cerebral ganglion; CP = cerebropleural ganglion; CPC = cerebropedal connective; CPL = cerebropleural connective; P = pedal ganglion; PL = pleural ganglion; POC = posterior connective; PPC = pleuropedal connective; VE = visceral loop element.

sumably homologous with the duct linking the statocyst and nerve in other adult pulmonates (KERKUT & WALKER, 1975). The formation of a single statolith in the larval statocyst and its replacement by statoconia during subsequent development (as in *Ovatella*) has been reported for other gastropods as well (e. g. *Haliotis*: CROFTS, 1937; *Onchidella*: FRETTER, 1943), suggesting that statoliths represent the more primitive evolutionary condition in gastropods.

The innervation mode of statocysts in adult gastropods is enigmatic. They are intimately connected with the pedal ganglia but innervated from the distant cerebral ganglia. The developmental sequence of the respective structures provides a possible explanation. As in *Ovatella*, the statocyst formation generally precedes that of the pedal ganglion (MOOR, 1983). Therefore, the statocysts must be initially innervated from the cerebral ganglia, a condition which is retained throughout development. The intimate connection of the statocyst nerve and the pleuropedal connective near the statocyst apparently represents a remainder of the larval organisation (where the pleuropedal connective and the statocyst nerve emerge from the same nerve trunk).

The transitory osphradium of *O. myosotis* presumably represents a phylogenetic recapitulation of this structure which is lacking in most adult non-aquatic pulmonates. A similar temporary osphradium has been reported in developing stylommatophorans (PELSENEER, 1901: *Helix aspersa*; HENCHMAN, 1890: *Limax maximus*).

The lack of the cephalic sensory organ in *Ovatella* (see formation of the cerebral commissure) is apparently due to the intracapsular development of this species: this organ may be reduced because it has no function during the larval phase inside the egg capsule. The planktotrophic larva of the ellobiid *Laemodonta octanfracta*, on the other hand, bears a well developed cephalic sensory organ with internally located ciliary bundles (pers. obs.). The presence of such an organ in the planktotrophic larva of another pulmonate, the onchidiid *Onchidium branchiferum* (pers. obs.), provides evidence that this is basic pattern in pulmonates. The connection of the cerebral commissure to the epithelium, which persists quite long in *Ovatella*, may be interpreted as a remnant of the cephalic sensory organ.

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